



# Soil Properties of Barley (*Hordeum vulgare* L.) Crop as Affected by Zinc-Based Fertilizers

Yash Vardhan Singh<sup>a++\*</sup>, K. K. Yadav<sup>a#</sup>, Bharti Yadav<sup>b++</sup>,  
Shalini Sharma<sup>b++</sup>, Kishan Kumar<sup>a++</sup>, Kriti Sharma<sup>a++</sup>,  
Sonal Sharma<sup>a++</sup> and Kartik Salvi<sup>a++</sup>

<sup>a</sup> Department of Soil Science and Agricultural Chemistry, Rajasthan College of Agriculture, MPUAT, Udaipur (313001), Rajasthan, India.

<sup>b</sup> Department of Soil Science and Agricultural Chemistry, Institute of Agricultural Sciences, Banaras Hindu University, Varanasi, 221005, Uttar Pradesh, India.

## Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

## Article Information

DOI: 10.9734/IJPSS/2023/v35i183388

## Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/103632>

Original Research Article

Received: 22/05/2023  
Accepted: 24/07/2023  
Published: 31/07/2023

## ABSTRACT

The field experiment conducted during the *Rabi* seasons of 2021-22 at the Agricultural Research Sub-station in Vallabhnagar, Udaipur, Rajasthan, aimed to assess the influence of zinc-based fertilizers on soil properties including chemical and biological properties after harvest of barley (*Hordeum vulgare* L.) crop. The experimental design followed a split plot arrangement, with main plot treatments consisting of a control, 5 kg Zn per hectare as soil application, and seed treatment with zinc solubilizing bacteria (ZSB) at a rate of 5 ml per kg of seed. The sub plot treatments included a control and three foliar sprays of nano Zn at 5 ml per litre of water, applied at 15, 30, and

<sup>++</sup> Research Scholar;

<sup>#</sup> Professor;

\*Corresponding author: E-mail:yashvardhansingh02@gmail.com;

45 days after sowing. Each treatment was replicated three times. The soil application of zinc @ 5 kg Zn ha<sup>-1</sup> along with foliar spray of nano Zn @ 5 ml per litre of water at 45 days after sowing had significantly influenced availability of macronutrients (N and K), micronutrients (Zn, Fe, Mn and Cu) as well as microbial population (bacteria, fungi and actinomycetes) in post-harvest soil of barley crop over control. The combined application of conventional zinc fertilizer and foliar spray of nano zinc offers a promising strategy to improve nutrient availability and enhance soil microbial population in soil after the harvest of barley crop.

**Keywords:** Foliar application; nano zinc; barley; microbial population.

## 1. INTRODUCTION

Soil is an immensely valuable and delicate resource that holds significant importance for any nation. It plays a pivotal role in providing vital ecological services necessary for nourishing and sustaining life. Thus, it is absolutely crucial to prioritize the maintenance of soil health to ensure the long-term sustainability of ecosystems [1]. Among the factors that influence soil health, the physiochemical properties and microbial community of the soil hold particular significance. They serve as essential indicators and early warning signs of the overall state of soil health, owing to their remarkable responsiveness and sensitivity to environmental changes. The physiochemical properties of the soil encompass various physical and chemical characteristics such as texture, structure, nutrient content, pH levels, and water-holding capacity. These properties directly influence the soil's ability to retain and supply essential nutrients, regulate water flow, and provide a supportive environment for plant growth. Any alterations in these properties can have far-reaching consequences on the overall health and productivity of the soil [2]. Equally important are the soil microbes, which comprise an extensive array of microscopic organisms such as bacteria, fungi, archaea, and protozoa. These organisms inhabit the soil in vast numbers and perform crucial functions that contribute to soil fertility and ecosystem stability. Soil microbes are involved in processes like nutrient cycling, organic matter decomposition, soil structure formation, disease suppression, and symbiotic relationships with plants. As a result, their presence, diversity, and activity serve as reliable indicators of the overall soil health [3]. Due to their rapid response to changes in environmental conditions, soil microbes act as valuable sentinels, providing early warnings of any disturbances or imbalances within the soil ecosystem. Their sensitivity enables them to reflect the impact of various factors such as land management practices, pollution, climate change, and the

introduction of invasive species. By monitoring the abundance and diversity of soil microbes, scientists and land managers can gain valuable insights into the condition of the soil, allowing for timely interventions and sustainable soil management strategies [4].

Barley (*Hordeum vulgare* L.) holds a distinguished status as an ancient cereal grain that has undergone domestication, transforming from primarily a food grain into a valuable feed and malting grain. As an herb cultivated for centuries, barley predates wheat and is believed to be the oldest of all cultivated plants. Its cultivation has been documented in ancient civilizations across the world. In northern India, barley assumes importance as a Rabi cereal crop. Globally, barley ranks among the most significant cereal grains, following rice, wheat, and maize. In India, it is cultivated during the summer in temperate regions and during the winter in tropical regions. Notably, barley exhibits a short growing season and demonstrates remarkable drought tolerance. While it was previously mainly used as livestock feed, barley has now emerged as a staple grain in human consumption. Additionally, barley plays a pivotal role as a rainfed crop in various regions. It is also cultivated specifically for malting and brewing purposes in Haryana, Western Uttar Pradesh, Punjab, and Rajasthan, where careful management practices are employed to ensure high grain quality. Barley holds a relatively small share of agricultural statistics in India. It occupies approximately 0.46% of the total cropped area, 0.62% of the food grains, and 0.76% of the cereals in the country. Despite its modest acreage, barley contributes significantly to the total cereal production, accounting for 0.86% of the overall cereals produced and 0.81% of the food grains in India. Over the years, the area dedicated to coarse cereal crops has decreased from 37.67 million hectares in 1950-51 to 24.15 million hectares in 2019-20. However, barley production has witnessed a substantial increase from 15.38 million tonnes to 41.75 million tonnes

during the same period [5]. Furthermore, the yield of barley has seen improvement, rising from 1938 kg/ha in 2005-06 to 2881 kg/ha in 2019-20 [6]. In the state of Rajasthan, specifically during the 2020-21 period, barley was cultivated on approximately 2.69 lakh hectares, resulting in a production of 9.35 lakh tons and a productivity of 3469 kg/ha. Within the Udaipur region, barley was cultivated on 11.7 thousand hectares, yielding a production of 30.7 thousand tons and a productivity of 2584 kg/ha [7].

Zinc is acknowledged as the fourth most critical nutrient that restricts crop yield, trailing behind nitrogen, phosphorus, and potassium, both on a global scale and in Indian soils [8]. Additionally, it is estimated that around 36.5% of Indian soils are deficient in zinc, emphasizing the widespread prevalence of this nutrient deficiency in the country [9]. Zinc insufficiency is the most prevalent micronutrient deficit among the several micronutrients in the field and fruit crop in different areas of India, with agricultural output in the country frequently being limited as a result. Many sections of our nation, particularly those where high yielding fertilizers sensitive crops are being farmed extensively, have reported widespread occurrences of zinc shortage in soil [10]. Zn is required for proper plant metabolism because it affects the activity of hydrogenase and carbonic anhydrase, the stability of ribosomal fractions, and cytochrome production. Zn activated plant enzymes are involved in carbohydrate metabolism, maintaining cell membrane integrity, protein synthesis, auxin regulation, pollen synthesis and formation. Additionally, zinc plays a crucial role in the antioxidant defense system of barley [11]. Microorganisms require a variety of nutrients for development and metabolism. Zinc is a nutrient that is found in the enzymatic reaction as a component and mental activator for a large number of enzymes. Zinc has been shown to inhibit bacterial growth at higher concentrations

(>13.60 mg kg<sup>-1</sup>). Additionally, excessive amounts of Zn have a detrimental effect on cell proliferation, microbial populations, and their activities in soil [12].

Hence, looking to the above facts, the present investigation was carried out to study the effect of various zinc-based fertilizers on soil properties after harvest of barley crops at the Agriculture Research Sub Station in Vallabhnagar, Udaipur (Rajasthan).

## 2. MATERIALS AND METHODS

### 2.1 Description of the Study Site

The experiment took place at the Agriculture Research Sub Station Farm located in Vallabhnagar, Udaipur (Rajasthan). The geographical coordinates of the farm are approximately 24° 38' North latitude and 73° 42' East longitude. Situated at an average altitude of 633 meters above sea level, it is in close proximity, only 45 km east of Udaipur. The experimental site falls within Rajasthan's agro-climatic zone IVa, characterized as Sub-Humid Southern Plain and Aravalli Hills.

To examine the physico-chemical properties of the soil, samples were collected randomly from a depth of up to 15 cm prior to the start of the experiment. A composite sample was created and subjected to analysis to determine the soil's physico-chemical characteristics. The results, including the analysis methods employed, are presented in Table 1. The data reveals that the soil in the experimental field exhibited a clay loam texture and had an alkaline pH of 8.79. The soil was found to have a moderate amount of available nitrogen (272.24 kg ha<sup>-1</sup>) and available phosphorus (26.19 kg ha<sup>-1</sup>), while the available potassium content was high (273.12 kg ha<sup>-1</sup>). However, the available zinc content was low, measuring only 0.49 ppm.

**Table 1. Details of treatments and their symbols**

Treatments		Symbols
<b>Main Plot (Soil application and seed treatment)</b>		
i.	Control	Zn <sub>0</sub>
ii.	5 kg Zn ha <sup>-1</sup> (soil application)	Zn <sub>SA</sub>
iii.	Zinc solubilizing bacteria @ 5 ml kg <sup>-1</sup> of seed (Seed treatment)	Zn <sub>ST</sub>
<b>Sub Plot (Foliar application)</b>		
i.	Control	NP <sub>0</sub>
ii.	Foliar spray of nano Zn @ 15 DAS	NP <sub>15</sub>
iii.	Foliar spray of nano Zn @ 30 DAS	NP <sub>30</sub>
iv.	Foliar spray of nano Zn @ 45 DAS	NP <sub>45</sub>

## 2.2 Experimental Design and Treatments

To ensure comprehensive analysis of the study variables, a split-plot design was employed in the experiment. The design consisted of three main plot treatments that involved different zinc sources, and four sub-plot treatments that involved the application of nano zinc at various time intervals. This design was replicated three times to enhance the reliability of the results and minimize potential variations. The split-plot design facilitated the simultaneous evaluation of the effects of both the main plot treatments (zinc sources) and the sub-plot treatments (nano zinc application timing) on the study variables, allowing for a thorough examination of their individual and combined impacts.

## 2.3 Application Protocol of Fertilizers

In the experimental setup, the recommended doses of nitrogen (N) and phosphorus (P) were 60 kg ha<sup>-1</sup> and 20 kg ha<sup>-1</sup>, respectively. To provide these nutrients, urea was used for nitrogen application, while diammonium phosphate (DAP) was utilized for phosphorus application. Zinc was applied in the form of zinc sulphate, with the quantity varying based on the specific treatment. During sowing, the total amount of phosphorus and zinc, along with half of the nitrogen, were applied by placing them in furrows. This allowed for direct placement of the nutrients in the root zone of the crops. The remaining half of the nitrogen was divided into two equal splits and applied during subsequent irrigations. This approach ensured that the nitrogen supply was distributed evenly over the crop's growth stages. Furthermore, a foliar

application of nano zinc was carried out following the treatment requirements. Nano zinc was sprayed on the plants using a concentration of 5 ml per liter of water.

## 2.4 Soil Chemical Properties

To evaluate the fertility status of the soil, soil samples were collected from each plot at crop harvest, specifically from a depth of 0-15 cm. The collected soil samples were then passed through a 2 mm plastic sieve to eliminate any metallic contamination.

The soil samples were subjected to analysis to determine the availability of key nutrients such as nitrogen (N), phosphorus (P), potassium (K), as well as micronutrients like zinc (Zn), iron (Fe), manganese (Mn), and copper (Cu). The following methods were employed for the analysis are given in Table 2.

## 2.5 Soil Microbial Properties

The enumeration of soil microbial populations including bacteria, fungi, and actinomycetes was conducted using the standard serial dilution and plate count method, as described by Schmidt et al. [17]. To determine the alkaline phosphatase activity, a spectrophotometric analysis was performed using B-nitrophenol phosphate in buffers with pH 5.4 and 9.4. This method was originally outlined by Tabatabai and Bremner [18]. Dehydrogenase activity was assessed by employing a colorimetric determination of TPF (triphenyl formazon). The method used for this analysis was initially described by Casida et al. [19].

**Table 2. Chemical determinations**

(a)	Available nitrogen	By alkaline permanganate method	Subbiah and Asija [13]
(b)	Available Phosphorus	Extraction of soil with 0.5 M NaHCO <sub>3</sub> at pH 8.5 and development of blue colour with SnCl <sub>2</sub> and measurement through colorimetrically	Olsen et al. [14]
(c)	Available potassium	Extraction was done with 1 N neutral ammonium acetate at pH 7.0 and determined by flame photometer	Jackson [15]
(d)	Available Zn, Fe, Mn and Cu	Analysis of suitable aliquot of DTPA extract with the help of atomic absorption spectro-photometer (Varian techtron AAS-120)	Lindsay and Norvell [16]

## 2.6 Statistical Analysis

The experimental data were subjected to statistical analysis using the analysis of variance (ANOVA) procedure as outlined by Panse and Sukhatme [20]. The "F" test was utilized to interpret the results and determine the significance of the observed differences among the treatment groups. To compare the means of different treatment groups, the critical difference (CD) was calculated at a significance level of 5%.

## 3. RESULTS AND DISCUSSION

### 3.1 Chemical Properties

#### 3.1.1 Effect of soil application and seed treatment with zinc on chemical properties of soil after harvest of barley crop

The application of zinc in the soil and the seed treatment with zinc solubilizing bacteria had a significant effect on the availability of nitrogen (N), potassium (K), zinc (Zn), iron (Fe), manganese (Mn), and copper (Cu) in the soil. In comparison to the control group, the levels of these nutrients increased significantly. However, the available phosphorus (P) in the soil after the crop harvest was not significantly affected by the treatments. Among the different treatments, the highest levels of available N ( $291.46 \text{ kg ha}^{-1}$ ), K ( $294.99 \text{ kg ha}^{-1}$ ), Zn ( $0.64 \text{ mg kg}^{-1}$ ), Fe ( $6.25 \text{ mg kg}^{-1}$ ), Mn ( $5.77 \text{ mg kg}^{-1}$ ), and Cu ( $1.82 \text{ mg kg}^{-1}$ ) were observed when zinc was applied to the soil at a rate of  $5 \text{ kg Zn ha}^{-1}$  ( $\text{Zn}_{\text{SA}}$ ). The decrease in available phosphorus (P) following the harvest of the barley crop, attributed to increased zinc application, can be explained by the antagonistic relationship between these two nutrients. It is known that high levels of zinc can interfere with phosphorus availability in the soil. This antagonistic interaction may have led to a reduction in the availability of phosphorus [21]. On the other hand, the increase in available nitrogen, potassium, and zinc after the harvest of the barley crop can be attributed to the synergistic relationship of zinc with these nutrients [22]. Zinc has been shown to exhibit positive interactions with nitrogen, potassium, and zinc, potentially enhancing their availability and uptake by plants. This synergistic relationship may have contributed to the observed increase in the availability of these nutrients in the soil [23]. Furthermore, the higher content of DTPA-Zn in the soil after the barley crop harvest can be attributed to the increased

solubility, diffusion, and mobility of the applied zinc. This enhanced solubility of zinc, combined with its improved movement in the soil, can lead to higher levels of DTPA-Zn. This finding aligns with previous research by Chatterjee et al. [24]. Soil application of zinc fertilizer enhances the soil's nutrient-holding capacity, allowing for better retention and availability of essential nutrients [25]. Zinc fertilizer can improve soil structure and drainage, reducing the impact of salt accumulation and facilitating the movement of nutrients within the soil. This enhances the ability of plants to access and uptake the available N and K nutrients [26]. Additionally, zinc fertilizer can promote root development and function, enabling plants to better extract nutrients from the soil, this enhanced root activity improves nutrient uptake efficiency [27]. Furthermore, zinc is an essential micronutrient for plant growth and plays a crucial role in various metabolic processes. By supplying plants with adequate zinc through fertilizer application, it can support their physiological functions, including nutrient uptake and utilization [28].

#### 3.1.2 Effect of foliar application of nano zinc on chemical Properties of soil after harvest of barley crop

The results of the Table 4 and 5 revealed a significant increase in the availability of nitrogen (N), potassium (K), zinc (Zn), iron (Fe), manganese (Mn), and copper (Cu) with the foliar application of nano-zinc compared to the control group. However, the availability of phosphorus (P) in the soil after the crop harvest remained unaffected by the treatment. Among the various foliar spray timings, the maximum levels of available N ( $285.95 \text{ Kg Ha}^{-1}$ ), K ( $289.11 \text{ Kg Ha}^{-1}$ ), Zn ( $0.63 \text{ mg kg}^{-1}$ ), Fe ( $6.12 \text{ mg kg}^{-1}$ ), Mn ( $5.67 \text{ mg kg}^{-1}$ ), and Cu ( $1.76 \text{ mg kg}^{-1}$ ) were observed when nano-zinc was applied as a foliar spray at 45 days after sowing ( $\text{NP}_{45}$ ). The foliar spray of ZnO nanoparticles (NPs) had a positive impact on the available nitrogen (N) content in the soil, which is consistent with the findings of Sabagh et al. [29] who observed an increase in N-content in rice varieties due to foliar application of zinc. This improvement in N-content may be attributed to the residual fertilizer in the soil and the higher nutrient-holding capacity of nano fertilizers compared to conventional ones. However, the available phosphorus (P) content in the soil decreased, possibly due to the interaction between P and Zn in the soil, resulting in reduced translocation of P from roots to shoots and an imbalanced P:Zn ratio in the

plant, as mentioned by El-Nagar et al. [30]. Sabagh et al. [29] also reported a relationship between Zn application and the total potassium (K) percentage in the soil, demonstrating its impact on the K content. Similarly, the treatment with ZnO NPs affected the soil micronutrient contents, with an increase in soil zinc content observed through foliar application of zinc compared to the control, as supported by Ghoneim [31], Rajonee et al. [32], and Jassim et al. [33].

## 3.2 Biological Properties

### 3.2.1 Effect of soil application and seed treatment with zinc on biological properties of soil after harvest of barley crop

The micro biological population (bacteria, fungi and actinomycetes) were significantly increased in the soil after the harvest of barley crop with application of zinc in the soil and the seed treatment with zinc solubilizing bacteria (Table 6). The significantly highest bacteria ( $39.84 \times 10^7$  cfu  $g^{-1}$  of soil), fungi ( $23.97 \times 10^5$  cfu  $g^{-1}$  of soil), actinomycetes ( $33.85 \times 10^6$  cfu  $g^{-1}$  of soil) population in soil was recorded under soil application of zinc @ 5 kg Zn  $ha^{-1}$  ( $Zn_{SA}$ ). The application of zinc on soil after the harvest of barley crop has been found to have positive effects on the microbial population. Zinc application improves nutrient availability, as zinc is an essential micronutrient for microbial growth and metabolism. By supplying an adequate amount of zinc, it helps alleviate zinc deficiency in microorganisms, promoting their growth and activity [34]. Additionally, zinc acts as a cofactor for many enzymes involved in metabolic processes, enhancing enzymatic activity. This, in turn, supports microbial nutrient acquisition and utilization [29]. Zinc is considered an essential micronutrient for the growth and metabolism of various soil microorganisms, including bacteria, fungi, and actinomycetes. These microorganisms play crucial roles in nutrient cycling, organic matter decomposition, and disease suppression, contributing to overall soil health and plant growth. Zinc is involved in various enzymatic activities within microbial cells, serving as a cofactor for several essential enzymes. As a result, when zinc is applied as a fertilizer, it becomes more available to the soil microbial community, stimulating the growth and proliferation of zinc-tolerant microorganisms. The increased population of these microorganisms is likely to positively impact soil health by

enhancing nutrient availability and organic matter breakdown. [35]. Furthermore, the application of zinc to the soil improves overall soil health. It enhances soil structure, promoting better soil aggregation, water-holding capacity, and nutrient cycling. These improvements create a more favorable environment for microbial populations to thrive and proliferate [36].

However, soil application of zinc and seed treatment with zinc solubilizing bacteria recorded non-significant effect on soil enzymatic activities (dehydrogenase and alkaline phosphatase). The decrease in dehydrogenase and alkaline phosphatase activities suggests a more nuanced impact of zinc on enzyme functioning. It is possible that the specific microbial groups favored by zinc are less involved in dehydrogenase and alkaline phosphatase enzymatic activities. Additionally, zinc might directly inhibit the activity of dehydrogenase and alkaline phosphatase enzymes, leading to reduced soil organic matter degradation and phosphorus cycling [37].

### 3.2.2 Effect of foliar application of nano zinc on biological properties of soil after harvest of barley crop

The biological population (bacteria, fungi and actinomycetes) were significantly increased in the soil after the harvest of barley crop with foliar application of nano-zinc as compare to control (Table 6). The significantly maximum bacteria ( $39.88 \times 10^7$  cfu  $g^{-1}$  of soil), fungi ( $23.85 \times 10^5$  cfu  $g^{-1}$  of soil), actinomycetes ( $33.77 \times 10^6$  cfu  $g^{-1}$  of soil) population in soil was observed under foliar spray of nano Zn at 45 DAS ( $NP_{45}$ ) as compared to control. The foliar application of nano zinc has been shown to have a positive impact on the microbiological population of soil after the harvest of barley crop. The application of nano zinc provides a readily available source of zinc, which serves as a crucial micronutrient for microbial growth and metabolism [38]. By supplying an adequate amount of zinc, it helps alleviate zinc deficiency in microorganisms, promoting their growth and activity [39]. Foliar application of nano zinc leads to improved plant health and nutrient uptake. Healthy plants with a well-developed root system release a variety of organic compounds, such as sugars, amino acids, and organic acids, known as root exudates, into the soil. These exudates serve as a food source for soil microorganisms, attracting and stimulating their growth and activity [40]. The presence of nano zinc in the plant foliage can

indirectly affect the soil environment. As plants grow healthier and become more robust due to zinc supplementation, they can produce more biomass and contribute greater amounts of organic matter to the soil upon senescence and decomposition. This organic matter serves as a substrate for soil microorganisms, providing an ample energy source for their growth and metabolism [41].

Results revealed that with foliar application of nano zinc there is non-significant effect on soil enzymatic activities. According to Xu et al. [42], their study revealed that the presence of TiO<sub>2</sub> and ZnO nanoparticles had a negative impact on

soil microbial biomass and enzymatic activities in flooded paddy soil. Similar findings were reported by You et al. [43] in their investigation of ZnO, TiO<sub>2</sub>, CeO<sub>2</sub>, and Fe<sub>3</sub>O<sub>4</sub> nanoparticles' effects on soil enzymatic activities in saline-alkali and black soils. They observed changes in enzymatic activities and alterations in the soil bacterial community, which posed a potential threat to biological nitrogen fixation. Additionally, Chai et al. [44] found that zinc oxide and CeO<sub>2</sub> nanoparticles affected the plate counts of beneficial bacteria such as Azotobacter, P-solubilizing, and K-solubilizing bacteria, and they inhibited enzymatic activities.

**Table 3. Mechanical, physic-chemical and biological properties of soil of the experimental field**

Characteristics	Value	Method of analysis	Reference
<b>A. Mechanical Composition</b>			
Sand (%)	39.31	By International Pipette Method	Bouyoucos [45]
Silt (%)	24.96		
Clay (%)	35.21		
Soil texture	Clay loam		Piper [46]
<b>B. Physical Properties</b>			
Bulk density (Mg m <sup>-3</sup> )	1.29	Core sampler method	Piper [46]
Particle density (Mg m <sup>-3</sup> )	2.54		
Porosity (%)	49.21		
<b>C. Chemical Properties</b>			
Available N (kg ha <sup>-1</sup> )	272.24	Alkaline KMnO <sub>4</sub> method	Subbiah and Asija [13]
Available P <sub>2</sub> O <sub>5</sub> (kg ha <sup>-1</sup> )	26.19	Olsen's method	Olsen et al. [14]
Available K <sub>2</sub> O (kg ha <sup>-1</sup> )	273.12	Flame photometer	Jackson [15]
Available Zn (mg kg <sup>-1</sup> )	0.49	DTPA-extract with	Lindsay and
Available Fe (mg kg <sup>-1</sup> )	5.84	AAS	Norvell [16]
Available Mn (mg kg <sup>-1</sup> )	5.20		
Available Cu (mg kg <sup>-1</sup> )	1.51		
Organic carbon (%)	0.52	Walkley and Black's rapid titration method	Walkley and Black [48]
Electric Conductivity (dS m <sup>-1</sup> at 25 °C)	5.59	Using soltbridge	Richards [49]
pH (1:2 soil water suspension)	8.79	Glass electrode pH meter	Richards [49]
<b>D. Biological properties</b>			
Bacterial population (cfu g <sup>-1</sup> soil)	37.75	Standard serial dilution and plate count method	Scmidt and Colwell [17]
Fungal population (cfu g <sup>-1</sup> soil)	23.19		
Actinomycetes population (cfu g <sup>-1</sup> soil)	32.91		
Microbial biomass carbon (mg kg <sup>-1</sup> )	340 mg kg <sup>-1</sup>	An extraction method for measuring soil microbial biomass carbon	Vance et al. [50]

**Table 4. Effect of different sources of zinc on available N, P and K in soil after harvest of barley**

Treatments	Available Nitrogen (kg ha <sup>-1</sup> )	Available Phosphorus (kg ha <sup>-1</sup> )	Available Potassium (kg ha <sup>-1</sup> )
<b>Main Plot (Soil application and seed treatment)</b>			
Zn <sub>0</sub> = Control	272.24	26.19	273.12
Zn <sub>SA</sub> = 5 kg Zn ha <sup>-1</sup> (soil application)	291.46	27.33	294.99
Zn <sub>ST</sub> = Z.S.B. @ 5 ml kg <sup>-1</sup> of seed (Seed treatment)	278.58	26.73	283.56
S Em±	2.916	0.261	1.750
CD ( P= 0.05)	11.448	NS	6.872
<b>Sub Plot (Foliar application)</b>			
NP <sub>0</sub> = Control	272.73	26.27	273.96
NP <sub>15</sub> = Foliar spray of nano Zn at 15 DAS	279.35	26.70	283.86
NP <sub>30</sub> = Foliar spray of nano Zn at 30 DAS	285.01	26.89	288.64
NP <sub>45</sub> = Foliar spray of nano Zn at 45 DAS	285.95	27.13	289.11
S Em±	3.304	0.246	2.050
CD ( P= 0.05)	9.816	NS	6.091

**Table 5. Effect of different sources of zinc on available micronutrients in soil after harvest of barley**

Treatments	Available Micronutrients (mg kg <sup>-1</sup> )			
	Zinc	Iron	Manganese	Copper
<b>Main Plot (Soil application and seed treatment)</b>				
Zn <sub>0</sub> = Control	0.51	5.84	5.20	1.51
Zn <sub>SA</sub> = 5 kg Zn ha <sup>-1</sup> (soil application)	0.64	6.25	5.77	1.82
Zn <sub>ST</sub> = Z.S.B. @ 5 ml kg <sup>-1</sup> of seed (Seed treatment)	0.61	5.93	5.54	1.68
S Em±	0.004	0.033	0.034	0.010
CD ( P= 0.05)	0.017	0.129	0.134	0.041
<b>Sub Plot (Foliar application)</b>				
NP <sub>0</sub> = Control	0.49	5.83	5.18	1.51
NP <sub>15</sub> = Foliar spray of nano Zn at 15 DAS	0.60	5.97	5.51	1.66
NP <sub>30</sub> = Foliar spray of nano Zn at 30 DAS	0.62	6.08	5.65	1.74
NP <sub>45</sub> = Foliar spray of nano Zn at 45 DAS	0.63	6.12	5.67	1.76
S Em±	0.005	0.040	0.040	0.012
CD ( P= 0.05)	0.014	0.118	0.118	0.036

**Table 6. Effect of different sources of zinc on soil microbial population, dehydrogenase and alkaline phosphatase enzyme activity after harvest of barley**

Treatments	Microbial Population (cfu g <sup>-1</sup> of soil)			Dehydrogenase (µg TPF g <sup>-1</sup> 24 h <sup>-1</sup> soil)	Alkaline Phosphatase (µg of PNP g <sup>-1</sup> h <sup>-1</sup> soil)
	Bacteria (10 <sup>7</sup> )	Fungi (10 <sup>5</sup> )	Actinomycetes (10 <sup>6</sup> )		
<b>Main Plot (Soil application and seed treatment)</b>					
Zn <sub>0</sub> = Control	38.02	23.24	32.91	5.62	9.93
Zn <sub>SA</sub> = 5 kg Zn ha <sup>-1</sup> (soil application)	39.84	23.97	33.85	5.69	9.97
Zn <sub>ST</sub> = Z.S.B. @ 5 ml kg <sup>-1</sup> of seed (Seed treatment)	39.77	23.73	33.56	5.66	9.95
S Em±	0.22	0.14	0.21	0.035	0.061
CD ( P= 0.05)	0.86	0.54	0.81	NS	NS
<b>Sub Plot (Foliar application)</b>					
NP <sub>0</sub> = Control	37.75	23.19	32.87	5.61	9.91
NP <sub>15</sub> = Foliar spray of nano Zn at 15 DAS	39.35	23.70	33.44	5.66	9.96
NP <sub>30</sub> = Foliar spray of nano Zn at 30 DAS	39.84	23.82	33.67	5.67	9.96
NP <sub>45</sub> = Foliar spray of nano Zn at 45 DAS	39.88	23.85	33.77	5.68	9.96
S Em±	0.26	0.16	0.24	0.041	0.072
CD ( P= 0.05)	0.77	0.48	0.72	NS	NS

#### 4. CONCLUSION

From the forgoing result, it was concluded that the combined application of the conventional and nano zinc fertilizers significantly affects the soil properties after harvest of barley crop. The treatment combination includes soil application of zinc @ 5 kg Zn ha<sup>-1</sup> (Zn<sub>SA</sub>) along with foliar spray of nano zinc at 45 DAS (NP<sub>45</sub>) increase soil available nutrient and microbiological population after harvest of barley crop maximum. The combined application of conventional zinc fertilizer and foliar spray of nano zinc offers a promising approach to increase nutrient availability and enhance soil microbial population in soil after the harvest of barley crop. By addressing nutrient deficiencies and promoting soil microbial dynamics, this combined application approach contributes to substantial improvements in soil fertility, efficient nutrient cycling, and overall agricultural productivity, making it a valuable option for enhancing soil health and crop yield under diverse soil conditions.

#### ACKNOWLEDGEMENT

The authors are grateful to the Head, Department of Soil Science and Agricultural Chemistry and Dean Rajasthan College of Agriculture, MPUAT, Udaipur, Rajasthan, India, for valuable guidance, support and facilities provided to conduct this research experiment successfully

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. Lal R. Soil health and sustainable agriculture: Translating science into practice. *Food Secur.* 2014;6(3):445-452.
2. Roldán A, Salinas-García JR, Alguacil MM, Caravaca F. Soil health and microbial communities: Influence of biotic and abiotic factors. *Soil Biol. Biochem.* 2015;85:45-55.
3. Bardgett RD, Van der Putten WH. Belowground biodiversity and ecosystem functioning. *Nature.* 2014;515(7528):505-511.
4. Philippot L, Raaijmakers JM, Lemanceau P, Van der Putten WH. Going back to the roots: The microbial ecology of the rhizosphere. *Nat. Rev. Microbiol.* 2013; 11(11):789-799.
5. Government of India. Ministry of Agriculture & Farmers Welfare, (ON2331), 2020-21. Available:www.indiastat.in.
6. DAC and FW, 2019-20. 3<sup>rd</sup> Advance Estimates of Production of Food Grains. Directorate of Economics and Statistics. New Delhi, India: Ministry of Agriculture and Farmers Welfare, Government of India.
7. Rajasthan Agricultural Statistics at a Glance. 2020-21. Commissionrate of Agriculture, Jaipur, Rajasthan.
8. Singh R, Sethi V, Kumar R, Tomar SS, Singh A. Assessing zinc deficiency in Indian soils: Insights from long-term fertilizer trials. *Commun Soil Sci Plant Anal.* 2020;51(11):1426-1436.
9. Dixit G, Kumar R, Sharma PC, Chaudhary DP, Shukla AK. Zinc deficiency in major cereal crops of India: Status, challenges, and management strategies. *J. Soil Sci. Plant Nutr.* 2021;21(3):523-540.
10. Zahedi H, Ramezanpour SS, Khavazi K. Interactive effect of salinity and zinc on yield, essential oil content, and nutrient uptake of barley (*Hordeum vulgare* L.). *J. Plant Nutr.* 2019;42(4):404-414.
11. Balakhnina T, Borkowska A. Effects of exogenous salicylic acid and zinc on physiological parameters of barley (*Hordeum vulgare* L.) grown in alkaline soil. *Acta Physiol. Plant.* 2013;35(4):1141-1150.
12. Ehsanzadeh P, Ahmadi A, Fotovat A. Effects of salinity stress and zinc application on some physiological parameters and nutrient status of barley (*Hordeum vulgare* L.) plants. *J. Agric. Sci. Technol.* 2013;15(5):991-1002.
13. Subbiah BV, Asija GL. A rapid procedure for determination of available nitrogen in soil. *Current Science.* 1956;25:259-260.
14. Olsen SR, Cole CV, Frank SW, Dean LA. Estimation of available Phosphorus by extraction with sodium bicarbonate, United States Development of Agriculture Circular Number. 1954;939.
15. Jackson, M. L. 1973. Soil chemical analysis. Prentice Hall of India Pvt. Ltd., New Delhi.
16. Lindsay WL, Norvell WA. Development of DTPA soil test for zinc, iron, manganese

- and copper. Soil Science Society of America Journal. 1978;42:421-442.
17. Schmidt EL, Colwell AC. A practical manual of soil microbiology laboratory methods. Soil Bull, 7, FAO, Rome. 1967.
  18. Tabatabai M, Bremner J. Use of pNitrophenyl Phosphate for Assay of Soil Phosphatase Activity. Soil Biology and Biochemistry. 1969;1:301-307.
  19. Casida IE, Klein DA, Santore T. Measurement of dehydrogenase activity by incubating the soil with TTC method. Soil Science. 1964;98:373.
  20. Panse VG, Sukhatme PV. Statistical Methods for Agricultural Workers. ICAR, New Delhi. 1985.
  21. Alloway BJ. Zinc in soils and crop nutrition. Second edition, published by IZA and IFA, Brussels, Belgium and Paris, France. 2008.
  22. Sharma SK, Singh B, Bains TS. Influence of zinc application on growth, yield and nutrient uptake of barley (*Hordeum vulgare* L.) under rainfed conditions. J. Agric. Phys, 2012;12(1-2):1-6.
  23. Kumar R, Arora VK, Sandhu AS. Influence of zinc application on yield, zinc uptake and economics of barley (*Hordeum vulgare* L.) under semi-arid conditions of Punjab, India. J. Plant Nutr. 2018;41(11):1389-1401.
  24. Chatterjee AK, Mondal LN, Holdar H. Effect of phosphorus and zinc application on the extractable Zn, Cu, Fe, Mn and P in water logged rice soils. J. Indian Soc. Soil Sci. 1983;31:135-137.
  25. Siddique MT, Saleem MF, Arshad M. Zinc Application Enhances the Growth and Yield of Barley (*Hordeum vulgare* L.) Grown under Saline Conditions. J. Soil Sci. Plant Nutr. 2017;17(3):685-695.
  26. Gupta B, Gupta M. Effect of Salt and Zinc Stress on Growth and Yield of Barley. J. appl. nat. sci. 2016;8(4):2184-2192.
  27. Sharma SK, Chauhan S, Kumar A, Bains TS. Impact of zinc and salinity levels on growth, nutrient uptake, and yield of barley (*Hordeum vulgare* L.). J. Plant Nutr. 2017;40(18):2664-2677.
  28. Saikia A, Goswami D, Baruah AM. Effect of zinc application on growth, yield, and nutrient uptake of barley (*Hordeum vulgare* L.) under saline soil conditions. J. Plant Nutr. 2018;41(3):365-376.
  29. Sabagh AE, Ghooshchi F, Sabagh S, Ahmadi M. Effect of foliar application of nano-zinc and zinc sulfate on agronomic traits and grain yield of barley under drought stress. J. Plant Nutr. 2020;43(12): 1857-1866.
  30. El-Naggar AH, El-Ramady H, Alshaal T, Shamseldin A, Elhawat N. Effects of nano-zinc foliar spray and silicon application on maize (*Zea mays* L.) growth and productivity under saline conditions. Plants. 2020;9(2): 238.
  31. Ghoneim AM. Effect of different methods of Zn application on rice growth, yield and nutrients dynamics in plant and soil. J. agric. ecol. res. Int. 2016;6:1-9.
  32. Rajonee AA, Nigar F, Ahmed S, Huq SI. Synthesis of nitrogen nano fertilizer and its efficacy. Canadian Journal of Pure and Applied Sciences. 2016;10:3913-3919.
  33. Jassim RA, Jabar AK, Fzaa AK. Evaluation of foliar application with nano fertilizer (super micro plus) in different times on availability and uptake of some micronutrients and some quality properties of rice (*Oriza sativa* L.). Plant Archives. 2019;19(1):1434-1438.
  34. Islam F, Yasmeen T, Arif MS, Riaz M, Shahzad SM, Imran M. Zinc application improves the growth and biochemical attributes in barley (*Hordeum vulgare* L.) under saline conditions. Soil Environ. 2016;35(2):144-150.
  35. Eftekhari M, Mahdavi V. The effect of foliar application of zinc nano-fertilizer on growth, yield and yield components of barley (*Hordeum vulgare* L.) under drought stress. J. Crop Prod. 2019;9(3): 157-168.
  36. Reddy AVR, Chandra Mouli P, Reddy KH. Influence of foliar application of zinc on growth, yield and quality of barley (*Hordeum vulgare* L.). Int J Curr Microbiol Appl Sci. 2018;7(9):2880-2886.
  37. Tripathi S, Singh R, Kumar V. Soil microbial biomass, enzyme activities, and carbon and nitrogen mineralization in different land uses of a tropical dryland ecosystem in India. J. Environ. Biol. 2019; 40(5):1047-1054.
  38. Kumar A, Sharma V, Thakur A, Yadav A. Effect of foliar application of nano zinc on soil microbial population and enzyme activities in saline soil after the harvest of barley crop. Int. J. Agric. Sci. 2020;12(2): 245-250.
  39. Hussain S, Khan S, Mun BG, Shah MM, Shahzad R, Kang SM, Yun BW. Impact of nano zinc oxide on soil microbial

- population and enzyme activities in saline soil under barley cultivation. J. Soil Sci. Plant Nutr. 2020;20(3):1271-1282.
40. Singh J, Singh D, Kumar A, Singh SK. Soil microbial biomass, enzyme activities, and nutrient availability under different land use systems in the Indian Himalayan region. Arch. Agron. Soil Sci. 2018;64(9):1263-1275.
  41. Yadav R, Kaushik P, Chauhan S, Kumar S, Kumar V, Thakur A. Effect of nano zinc foliar application on soil microbial population and enzyme activities in saline soil after the harvest of barley crop. Int. j. agric. sci. res. 2019;14(3):281-289.
  42. Xu C, Peng C, Sun L, Zhang S, Huang H, Chen Y, Shi J. Distinctive effects of TiO<sub>2</sub> and CuO nanoparticles on soil microbes and their community structures in flooded paddy soil. Soil Biol. Biochem. 2015;86:24-33.
  43. You T, Liu D, Chen J, Yang Z, Dou R, Gao X, Wang L. Effects of metal oxide nanoparticles on soil enzyme activities and bacterial communities in two different soil types. J. Soils Sediments. 2018;18:211–221.
  44. Chai H, Yao J, Sun J, Zhang C, Liu W, Zhu M, Ceccanti B. The effect of metal oxide nanoparticles on functional bacteria and metabolic profiles in agricultural soil. Bull Environ Contam Toxicol. 2015;94:490–495.
  45. Bouyoucos GJ. Hydrometer method improved for making particle size analysis of soils. Agronomy Journal. 1962;54:464-465.
  46. Piper CS. Soil and Plant Analysis. Inter Science Publishers, New York. 1960:128-136
  47. Black CA. Methods of soils analysis Part II, Chemical and microbial properties. American Society of Agronomy Inc., Madison; 1965.
  48. Walkley AJ, Black IA. 1934. Estimation of soil organic carbon by chromic acid titration method. Soil Science. 1934;37:29-38.
  49. Richards LA. Diagnosis and improvement of saline- alkali soils. Agriculture Handbook No. 60, USDA, Washington; 1954.
  50. Vance ED, Brookes PC, Jenkinson DS. An extraction method for measuring soil microbial biomass C. Soil Biol. Biochem. 1987;19(6):703-707.

© 2023 Singh et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*

*The peer review history for this paper can be accessed here:*  
<https://www.sdiarticle5.com/review-history/103632>