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# Therapeutics Activities of Amazonian Plant Himatanthus sucuuba (Spruce ex Müll. Arg.) Woodson (Apocynaceae): A Review

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#### Authors' contributions

This work was carried out in collaboration among all authors. The authors equally worked during elaboration of manuscript, as in conceptualization, and writing (original draft preparation, review and editing). All authors read and approved the final manuscript.

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**Review Article** 

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## ABSTRACT

Several compounds extracted from medicinal plants and their active ingredients can relieve symptoms and even cure diseases, although they occasionally have adverse effects. The knowledge of their properties has been transmitted over the centuries within and among human communities. *Himatanthus sucuuba* is an Amazon plant that has its value attributed to the different herbal impacts popularly reported. This review presents significant biological activities such as antibacterial, antifungal, anthelmintic, antileishmanial, anti-inflammatory, analgesic, antidepressant, immunoregulatory, cytotoxic, and genotoxic. Thus, provides a synopsis of the literature exploring the extracts of *H. sucuuba* to the Amazon region that could efficiently prevent pathologies associated with leishmaniosis, infection for bacteria or fungus, depression, oxidative stress, and cytotoxicity or genotoxicity.

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#### 1. INTRODUCTION

#### **1.1 Phytotherapy: Historical Aspects**

Ancient people, such as Egyptians, Greeks, Hindus, and Persians, explored plant species for therapeutic purposes, seeking alternatives for the treatment of their pathologies, such knowledge being transmitted to later civilizations, thus generating, through observation, empirical knowledge, giving rise to phytotherapy [1,2]. The first records on the use of medicinal plants are dated to 500 BC, in the Chinese text that reports names, doses, and indications of plants' use to treat diseases [3]. One of the oldest records related to phytotherapy is the Egyptian papyrus Ebers dated in 1500 BC, and it is undoubtedly the oldest medical text recorded where it documents more than 800 herbal medicines and their actions in the treatment of common diseases at that time and which are still used today [4]. Other records also occurred as an example, in the period 2838-2698 BC, when the Chinese Emperor Shen Nung classified 365 poisons and medicinal herbs that were used under the Taoist inspiration of Pan Ku. considered God of creation [5].

In South America, indigenous ethnic groups have long used medicinal plants in rituals for healing and worshiping the shaman, an individual in whom he was responsible for conducting the ritualism, people who were found in countless tribes [6]. In Brazil, during the centuries of colonization, there was miscegenation of various cultures and empirical knowledge, such as the African peoples who consumed herbs and the indigenous people who used plants with potential for healing through the chief of the village and the European peoples when they arrived in Brazil, came across local knowledge [7].

## **1.2 Amazon Forest: Medicinal Plants**

The Amazon Forest is an ecosystem with incredible biodiversity, being part of this arsenal consisting of medicinal plants commonly consumed among traditional populations [8]. The establishment of a close relationship with the environment allowed resources to diverse peoples, called traditional people, such as the indigenous, caboclos, riverside, extractivist, and decent quilombola community [9].

Approximately 49.29% of the Brazilian territory, the Amazon is today the largest biome in the world, covering nine countries (Brazil, Paraguay, Bolivia, Peru, Ecuador, Colombia, Venezuela, French Guiana, and Suriname), with around 40 thousand species of plants, 300 species of mammals, 1,300 species of birds cataloged, which live in 4,196,943 km<sup>2</sup> of dense and open forests [10].

The vegetal diversity of the Amazon Forest corresponds to the arboreal analysis of several species to define which are native to the Amazon and which have been introduced over time [11,12]. The families with the highest number of Amazon representatives are Leguminosae, Orchidaceae, Melastomataceae, Rubiaceae. Araceae. Myrtaceae. Lauraceae. Annonaceae. Poaceae. and Euphorbiaceae. which corresponds to 55% of all species [12]. Among medicinal plants, we will highlight the Himathantus sucuuba (Apocynaceae), or sucuuba as it is popularly known, which has aroused great interest on the part of researchers due to the production of latex, which has antiinflammatory and analgesic action, healing effect, and antibacterial activity, in addition to its previously isolated chemical compounds, which proved activitv against nasopharvngeal carcinoma [13].

## **1.3 Apocynaceae Family**

The Apocynaceae family is widespread in tropical and subtropical regions [14], which can be considered one of the largest and most important plant sources of chemical constituents since alkaloids' production is present in laticiferous tissues. These characteristics are closely related linked to plant defense against herbivores. Still, for centuries it has been used by ethnic groups as a source of alternative medicines, and its use continues today [15]. The family plants are included phylogenetically in the order Gentiales and subclass Asteridae, having as a significant characteristic the presence of latex in non-articulated, branched, or unbranched laticiferous structures [16]. Until 2009, the family Apocynaceae was called Asclepiadaceae, and its change was mediated by the classification system Phylogeny Group III [17].

The Apocynacea family consists of approximately 355 genera and 3700 species but

95 genera and 850 species occur in Brazil [18]. Also, represented by shrubs, woody, herbaceous and ornamental plants, with major classes of metabolites as alkaloids, triterpenoids, iridoids, and cardenolides Pregnanes, flavonoids and phenolic acids represent the minor classes [19-21]. Studies suggest significant biological effects as antioxidant, antimicrobial, antiinflammatory, hepatoprotective and cytotoxic effects [21].

# 1.4 Genus *Himatanthus*

In 1819, the genus Himatanthus was described by Willdenow, being composed of species native to South America represented by trees, shrubs, lactescent sub-shrub [22]. Initially, or Himatanthus was included in the genus Plumeria; however, a decisive factor for the separation was the presence of large bracts surrounding the flower buds in Himatanthus, where their presence inspired the name of this genus, which means 'floral mantle' [23]. In the genus Himatanthus there are 13 species: H. articulatus (Vahl) Woodson, H. attenuatus (Benth.) Woodson, H. bracteatus (A. DC.) Woodson, H. drasticus (Mart.) Plumel, H. fallax (Muell. Arg.) Plumel, H. lancifolius (Muell. Arg.) Woodson, H. obovatus (Muell. Arg.) Woodson, H. obovatus (Muell. Arg.) Woodson, H. phagedaenicus (Mart.) Woodson, H. semilunatus Markgraf, H. speciosus (Muell. Arg.) Plumel, H. stenophyllus Plumel, H. sucuuba (Spruce) Woodson and H. tarapoteensis (Schumann ex Markgraf) Plumel [24]. The species of Himatanthus are trees with woody branches and may be minor at the beginning of their development and reach up to 20 to 30 meters in height, yet the species show early flowering [25].

The significant medicinal potential of the genus Himatanthus has been described in latex (hypoglycemic activity [26], antioxidant [27], genotoxicity [28], antiinflammatory [29], antitumor [30], and gastroprotective [31]) and leaves (antimicrobial [32]) from H. drasticus. Also, the latex of H. articulates represents therapeutic action to mutagenesis [33], antimicrobial [34], and antiproliferative activity [35]. However, the H. bracteatus bark showed important action on vascular and non-vascular smooth muscle [36], gastroprotective [37], acetylcholinesterase inhibition [38], anti-inflammatory effect [39], antimicrobial [40], and antiviral [41] activities. Among the species described, H. sucuuba stands out for presenting several therapeutic actions.

# 1.5 Himatanthus sucuuba

Himatanthus sucuuba (Spruce ex Müll. Arg.) Woodson, belonging to the Apocynaceae family, order Gentianales and subclass Asteridae is a large tree species, reaching 20 to 30 m in height, with elliptical ovate or oblong leaves (20-28 cm long and 5-7 cm wide), with acuminated or acute attenuated apex abruptly base and inflorescences with 1-1.15 cm long, white flowers, with jagged lobes cup [42]. It also colonizes the floodplain and rigid ground regions in the Central Amazon; its value is attributed to the different phytotherapeutic effects [43,44]. H. sucuuba, popularly known in the northern region of Brazil as sucuuba, janaguba or sucuba [45]. The fruits are elongated and green when immature, and dark brown when ripe (2.5 cm long and 3.5 cm in diameter) [46], having numerous dry ellipsoid seeds, surrounded by a circular membrane wing well developed, this structure being important in the dispersion of the species by the wind (Fig. 1) [23]. Still, its flowering occurs during a long period of the year, predominating in August to October, with the fruits ripening from March to May [47].

*H. sucuuba* is a latescent species, with an erect trunk and rough skin [48] popularly the use of latex is used against skin conditions, while the leaves are used in the form of decoction against constipation, pain and stomach irritation [49]. Studies indicate the use of latex and bark of the trunk of *H. sucuuba* showed anti-inflammatory and analgesic action, therefore, indicated for the treatment of arthritis, edema, leishmaniasis and intestinal diseases [50-52] (Table 1).

#### 2. PHYTOCHEMICAL COMPOSITION AND BIOLOGICAL ACTIVITIES OF Himatanthus sucuuba

This review presents the analgesic, antibacterial, antidepressive, antifungal, anthelmintic, antiinflammatory, immunoregulatory, antileishmanial, cytotoxic, genotoxicity, and teratogenic activities from *H. Sucuuba* extracts. Besides, *in vitro*, *in vivo*, and *ex vivo* experimental models are shown in Table 1.

#### 2.1 Analgesic

Pain is a physiological and pathological phenomenon; therefore, a symptom of various diseases [67] remains one of the most poorly understood sensations, with poor clinical management despite many pain-relieving drugs available on the market.

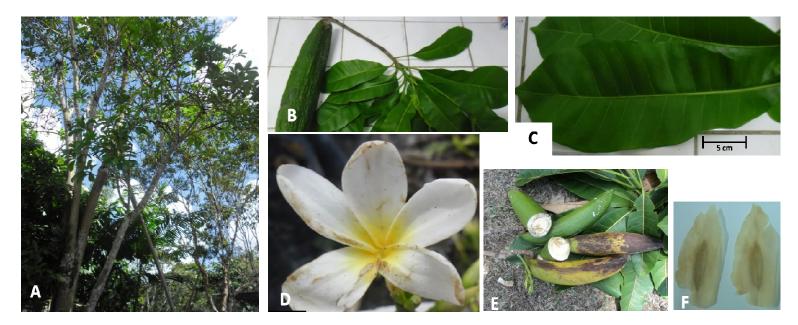


Fig. 1. Morphological aspects of the *Himatanthus sucuuba*. (A) The adult plant; (B) Immature fruit and branch; (C) Leaf detail; (D) Flower details; (E) Immature and mature fruits; (F) Seeds [16,52].

Biological compound	Vegetative Parts	Extraction	Therapeutic Activity	Concentration	Experimental model
*	B [53]	EtOH/H <sub>2</sub> O [53]	Skeletal anomalies in fetuses [53]	500 mg/kg (p.o.) [53]	<i>in vivo</i> : Female Wistar rats [53]
*	B [54]	*	Capillary permeability [54]	2.5 – 5.0 mg [54]	<i>in vivo</i> : Male albino rats [54]
*	B [55]	EtOH - Hex fraction [55]	Cytotoxic [55]	GI <sub>50</sub> 0.125 mg/mL [55]	<i>in vitro</i> : BALB/c 3T3 cell lines [55]
*	B [55]	EtOH - Hex fraction [55]	Cytotoxic [55]	GI <sub>50</sub> 0.080 mg/mL [55]	<i>in vitro</i> : H-460 cell lines [55]
*	B [56]	EtOH [56]	Antileishmanial [56]	IC <sub>50</sub> 0.9 μΜ [56]	in vivo: BALB/c mice [56]
*	L [49]	Hex [49]	Antiinflammatory [49]	200 mg/kg (p.o.) [49]	in vivo: rat [49]
*	L [57]	EtOH/H <sub>2</sub> O [57]	Analgesic [57]	1000 mg/kg (p.o.) and 100 mg/kg (i.p.) [57]	<i>in vivo</i> : Male swiss mice [57]
*	L [50]	n-ButOH [50]	Antileishmanial [50]	IC <sub>50</sub> 15.7 µg/mĹ [50]	in vivo: BALB/c mice [50]
*	L [58]	*	Anthelmintic [58]	3 mg/L [58]	in vivo: Humans [58]
*	L [59]	*	Genotoxicity [59]	0.01 – 0.03 g/mL [59]	<i>ex vivo</i> : Humans lymphocytes [59]
*	P [60]	EtOH [60]	Antibacterial [60]	37,17 mg [60]	in vitro: Clostridium histolyticum, and Bacteroides fragilis [60]
Cinnamates	L [49]	Hex [49]	Antiinflammatory [49]	100 mg/kg (p.o.) [49]	in vivo: rat [49]
Confluentic acid	B [61]	MeOH [61]	Antidepressive [61]	IC <sub>50</sub> 0.22 µM [61]	<i>in vivo</i> : Brains of male Wister rats [61]
Isoplumericin	B [62]	Hex [62]	Antifungal [62]	1 µg [62]	in vitro: Cladosporium sphaerospermum [62]
Lupeol acetate	R [63]	MeOH [63]	Immunoregulatory [63]	50 mg/mL [63]	ex vivo: Murine macrophages and splenocytes from BALB/c mice [63]
Lupeol cinnamate	R [63]	MeOH [63]	Immunoregulatory [63]	50 mg/mL [63]	ex vivo: Murine macrophages and splenocytes from BALB/c mice [63]

# Table 1. Biological compounds from *Himatanthus sucuuba* extracts

Biological compound	Vegetative Parts	Extraction	Therapeutic Activity	Concentration	Experimental model
Plumericin	B [62]	Hex [62]	Antifungal [62]	1 µg [62]	in vitro: Cladosporium sphaerospermum [62]
Plumericin	B [64]	*	Antiinflammatory [64]	3 mg/kg <sup>-1</sup> (i.p.) [64]	<i>in vivo</i> : C57BL/6J male mice [64]
Plumericin	R [65]	*	Antifungal [65]	2.5 µg [65]	in vitro: Saccharomyces cerevisae ATCC 2601 [65]
Plumericin	R [65]	*	Antifungal [65]	1.25 µg [65]	in vitro: Candida albicans ATCC 10231 [65]
Plumericin	R [65]	*	Antifungal [65]	2.5 µg [65]	in vitro: Candida dubliniensis SM26 [65]
Plumericin	R [63]	MeOH [63]	Immunoregulatory [63]	50 mg/mL [63]	<i>ex vivo</i> : Murine macrophages and splenocytes from BALB/c mice [63]
Plumericin and Isoplumericin	L [66]	Hex [66]	Antifungal [66]	IC <sub>12</sub> 112.3 µg/mL [66]	in vitro: Saccharomyces cerevisae [66]

Legend: \*, Not reported; B, bark, L, latex; P, plant; R, roots; EtOH/H<sub>2</sub>O, hydroethanolic; EtOH, ethanol; Hex, hexane; MeOH, methanol; n-ButOH, n-Butanol; GI<sub>50</sub>, growth inhibition in 50%; IC<sub>12</sub>, concentration required to produce a zone of inhibiton of 12 mm; IC<sub>50</sub>, inhibition of biological process or biological component by 50%; p.o, oral administration; i.p, intraperitoneal injection

Numerous studies have been conducted to elucidate neuronal pathways of pain and develop new treatments [68] with synthetic or natural compounds. Leaves EtOH/H<sub>2</sub>O extracts from *H. sucuuba* (1000 mg/kg (p.o.) and 100 mg/kg (i.p.)) showed excellent effect analgesia in male swiss mice [57] related with ethanolic extract from *Hydrocotyle umbellata* (4000 mg/kg (p.o.)) [69] or methanolic extract of *Listea glutinosa* leaves (500 mg/kg (i.p.)) [70].

# 2.2 Antibacterial

Antibacterials are used to minimize the toxic effects caused by bacteria, but the inappropriate use of such compounds promotes microbial resistance. Thus, actions must be taken to reduce this problem, such as controlling the use of antibiotics, developing research to understand the genetic mechanisms of resistance better, and continuing studies to develop new drugs, whether synthetic [71] or natural [72]. Ethanolic extract of H. sucuuba (37,17 mg, d>7 mm) inhibited Clostridium histolyticum, and Bacteroides fragilis [60]. However, ethanolic extracts of Punica granatum (15 mg/mL) significantly suppressed the development of Staphylococcus aureus (d>23.7 ± 0.35 mm), Pseudomonas aeruginosa (d>22.6 ± 0.74 mm). Also, S. aromaticum (15 mg/mL) was able to inhibited S. aureus (d>19.3 ± 0.65 mm), and P. aeruginosa (d>17.5 ± 0.35 mm) cultures [73].

## 2.3 Antidepressive

Disorders of the central nervous system, such as neurodegenerative diseases, are widely studied. Pathological brain damage mechanisms are associated with an inflammatory reaction, bloodbrain barrier rupture, oxidative stress, and neuronal apoptosis. Studies suggest that therapeutic effects with neuroprotective molecules against brain damage [74]. Confluentic acid from bark methanolic extract of H. sucuuba showed an essential inhibition of human monoamine oxidase (MAO) (IC<sub>50</sub> 0.22 µM) in male Wister rats' brains [61]. Demonstrating a significant therapeutic effect compared to kushenol F (IC\_{50} 69.9  $\mu M)$  and formononetin (IC<sub>50</sub> 13.2 µM) from methanolic root extract of Sophora flavecens [75] or extracts of Hypericum perforatum (IC<sub>50</sub> 0.134 mM) [76].

## 2.4 Antifungal

Fungi represent a food source, but some are responsible for infections, so synthetic or natural

antifungals are used. Some molecules extracted from barks (isoplumericin and plumericin), roots (plumericin), and leaves (plumericin and isoplumericin) of *H. sucuuba* showed inhibition in a different culture as *Cladosporium sphaerospermum* [62], *Saccharomyces cerevisae* ATCC 2601 [65], *Candida albicans* ATCC 10231 [65], *Candida dubliniensis* SM26 [65], and *Saccharomyces cerevisae* [66].

Studies with isoplumericin or plumericin () from bark hexane extract of *H. sucuuba* inhibited with *C. sphaerospermum* [62]. Plumericin from roots extract of *H. sucuuba* interfered with 2.5 µg the development *S. cerevisae* ATCC 2601 [65], 1.25 µg *C. albicans* ATCC 10231 [65], and 2.5 µg *C. dubliniensis* SM26 [65]. Also, plumericin and isoplumericin from leave extract of *H. sucuuba* (IC<sub>12</sub> 112.3 µg/mL) inhibited *S. cerevisae* [66]. Moderate antifungal activity compared to synthetic compounds [77].

# 2.5 Anthelmintic

Parasitic diseases are responsible for high morbidity and mortality in animals worldwide, considerable losses for food production, and promoting malnutrition in humans. Studies that plant extracts have an antiparasitic effect as a latex of *H. sucuuba* 3 mg/L in humans [58] or the whole plant extract (5 mg/mL) of *Picria felterrae Lour* against *Haemonchus contortus* [78] showed a significant anthelmintic action.

## 2.6 Antiinflammatory and Immunoregulatory

Natural products that carry biological activities are consumed daily to help maintain human health. The constant search for bioactive compounds with antiinflamatory and immunoregulatory actions is of interest to research centers [79]. Leave hexane extract 200 mg/kg (p.o.), Cinnamates leave hexane extract 100 mg/kg (p.o.) in rat [49], and Plumericin from bark extract 3 mg/kg-1 (i.p.) in C57BL/6J male mice [64] from *H. sucuuba* showed a moderate antiinflamatory and vascular permeability [54] effects related to synthetic compounds [80].

Various plant-based extracts have been shown to exert protective effects on the dysfunctional mechanisms involved in immunoregulatory integrity and function. These essential compounds are found in plants (*Ginkgo biloba*, *Panax ginseng*, and *Bacopa monnieri*) and food sources (Resveratrol, Curcumin, Pinocembrin, Epigallocatechin-3-gallate, Berberine and Caffeine) [81]. Also, Lupeol cinnamate and Plumericin methanol extracted from the root of *H. sucuuba* showed an immunoregulatory effect in murine macrophages and splenocytes from BALB/c mice [63].

# 2.7 Antileishmanial

Leishmaniasis is a disease with ample clinical spectrum and epidemiological diversity and is considered a sizable public health problem [82]. Several treatments, including plant extracts, showed an effective action against Leishmania as methanolic leave extract of *Casearia arborea* [83], ethanolic leave extract of *Ocimum sanctum* [84], hexane leave extract of *Croton caudatus* [85], ethanolic bark extract [56], and n-butanol leave extract [50] from *H. sucuuba*.

# 2.8 Cytotoxic and Genotoxicity

Several compounds can induce mechanisms on the cell cycle progression, therefore, acting as cytotoxic and genotoxicity. Ethanolic extract of seeds of *Euphorbia lathyrism* [86] and roots extract of *Glycyrrhiza iconica* [87] exhibited potent cytotoxicity against cancer cell lines. Also, ethanolic/hexane fractions from the bark [55] and leave [59] from *H. sucuuba* showed cytotoxicity and genotoxicity against BALB/c 3T3 cell lines [55], H-460 cell lines [55], and human lymphocytes [59].

## 2.9 Teratogenic

Congenital anomalies (teratogenic processes) are already the second cause of infant mortality in animals and result from genetic and environment factors, but a multifactorial etiology has been observed. Humans are exposed to millions of potentially harmful substances and hazardous conditions daily [88]. However, only a tiny part of these substances have been tested in animals. Even fewer were confirmed as teratogenic like extract of *Peumus boldus*, *Foeniculum vulgare*, *Melissa officinalis*, and *Mentha piperita L*. [89]. Hidroethanolic extract from roots of *H. sucuuba* showed skeletal anomalies in fetuses in Wistar rats [53].

## **3. CONCLUSION**

This review provides an overview of *H. sucuuba* from the Amazon region and some biological activities as antimalarial known by the local population. Also, recent studies with extracts

described significant as antibacterial, antifungal, anthelmintic, antileishmanial, anti-inflammatory, analgesic, antidepressant, immunoregulatory, cytotoxic, and genotoxic. However, recent studies would be considering the enormous potential of protecting against diseases associated with leishmaniosis, infection for bacteria or fungus, depression, oxidative stress, and cytotoxicity or genotoxicity.

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## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

# REFERENCES

 Salesse D, Medeiros FC, Silva CCM, Lourenço ELB, Jacomassi E. Etnobotânica e Etnofarmacologia das espécies de Amaryllidaceae, Anacardiaceae, Annonaceae e Apiaceae. Arquivos de Ciências da Saúde da UNIPAR. 2018; 22(3). Available:https://doi.org/10.25110/argsaud

Available:https://doi.org/10.25110/arqsaud e.v22i3.2018.6376

 Rocha FAG, Araújo MFF, Costa NDL, Silva RP. O uso terapêutico da flora na história mundial. HOLOS. 2015;1(31):49-61.

Available:https://doi.org/10.15628/holos.20 15.2492

- Duarte MCT. Atividade antimicrobiana de plantas medicinais e aromáticas utilizadas no Brasil. Revista MultiCiência. 2006;7. Available:http://www.multiciencia.unicamp. br/artigos\_0 7/a\_05\_7.pdf
- Saad MES. Plantas medicinales en el antiguo Egipto; 2015. Available:https://core.ac.uk/download/pdf/4 3551207.pdf
- Vale NB. A farmacobotânica, ainda tem lugar na moderna anestesiologia? Rev Bras Anestesiol. 2002;52(3):368-380. Available:https://www.scielo.br/pdf/rba/v52 n3/v52n3a13.pdf
- 6. Braga CM. Histórico da utilização de plantas medicinais; 2011.

Available:https://www.bdm.unb.br/bitstrea m/10483/1856/1/2011\_CarladeMoraisBrag a.pdf

 Tomazzoni MI, Negrelle RRB, Centa ML. Fitoterapia popular: A busca instrumental enquanto prática terapeuta. Texto Contexto–Enferm. 2006;15(1):115-121.

Available:http://dx.doi.org/10.1590/S0104-07072006000100014

 Barquete CC, Belfort AS, Santos JF, Souza SF. Fitoterápicos amazônicos:Copaíba no tratamento de feridas cutâneas. Enciclopédia Biosfera, Centro Científico Conhecer–Goiânia. 2017 ;14(26). Available:https://doi.org/10.18677/EnciBio\_

2017B32

 Castro E. Território, biodiversidade e saberes de populações tradicionais. In: Castro E, Pinton F. (Org.). Faces do trópico úmido: conceitos e novas questões sobre desenvolvimento e meio ambiente. Belém: CEJUP; UFPA-NAEA. 1997;221-242.

> Available:http://nupaub.fflch.usp.br/sites/nu paub.fflch.usp.br/files/Ecologia%20e%20C osmologia147.pdf

- ICMBio. Instituto Chico Mendes de Conservação da Biodiversidade. Amazônia; 2010. Available:https://www.icmbio.gov.br/portal/ unidadesdeconservacao/biomasbrasileiros/amazonia
- Steege HT, Oliveira SM, Pitman NCA, Sabatier D, Antonelli A, Andino JEG, et al. Towards a dynamic list of Amazonian tree species. Sci Rep. 2019;9:3501. Available:https://doi.org/10.1038/s41598-01940101-y
- Cardoso D, Särkinen T, Alexander S, Amorim AM, Bittrich V, Celis M, et al. Amazon plant diversity revealed by a taxonomically verified species list. PNAS. 2017;114(40):10695-10700.

Available:https://doi.org/10.1073/pnas.170 6756114

- Nascimento AP, Neto PQC, Pereira JO. Potencial biotecnológico de bactérias endofíticas de *Himatanthus sucuuba*; 2017. Available:https://tede.ufam.edu.br/handle/t ede/5842
- Bhadane BS, Patil MP, Maheshwar VL, Pati RH. Etnofarmacologia, fitoquímica e avanços biotecnológicos da família

Apocynaceae: Uma revisão. Phytother Res. 2018;32(7):1181-1210. Available:https://doi.org/10.32712/2446-4775.2019.800

- 15. Oliveira AA, Silva JRA. Análise fitoquímica dos extratos e frações obtidos de *Himatanthus sucuuba*; 2013. Available:https://tede.ufam.edu.br/handle/t ede/4498
- Pereira MM, Jácome RLRP, Alcantara AF, Alves RB, Raslan DS. Alcalóides indólicos isolados de espécies do gênero Aspidosperma (*Apocynaceae*). Quim Nova. 2007;30(4):970-983. Available:https://doi.org/10.1590/S0100-40422007000400037
- 17. Endress ME, Liede-Schumann S, Meve U. An updated classification for Apocynaceae. Phytotaxa. 2014;159(3):175 -194.

Available:http://dx.doi.org/10.11646/phytot axa.159.3.2

- Souza VC, Lorenzi HJ. Systematic Botany: Illustrated guide for identifying families of native and exotic phanerogams in Brazil, based on APG II. Nova Odessa: Plantarum Institute; 2008.
- Aiyambo D. Traditional uses of selected members of the *Apocynaceae* family in Namibia. Ministry of Agriculture, Water and Forestry, Windhock; 2010. Available:http://www.nbri.org.na/traditionaluses-of-selected-members-of-theapocynaceae-family-namibia
- Joselin J, Brintha TSS, Florence AR, Jeeva S. Screening of select ornamental flowers of the family Apocynaceae for phytochemical constituents. Asian Pac J Trop Dis. 2012;2:260-264. Available:https://doi.org/10.1016/S2222-1808(12)60162-5
- 21. Chan EWC, Wong SK, Chan HT. Apocynaceae species with antiproliferative and/or antiplasmodial properties: A review of ten genera. J Integr Med. 2016;14(4): 269-284.

Available:https://doi.org/10.1016/S2095-4964(16)60261-3

- Larrosa CRR, Duarte MR. Contribution to the anatomical study of the stem of *Himatanthus sucuuba* (Spruce ex Müll. Arg.) Woodson, *Apocynaceae*. Rev Bras Farmacogn. 2005;15(2):110-114. Available:https://doi.org/10.1590/S0102-695X2005000200007
- 23. Plumel MM. The genre *Himatanthus* (*Apocynaceae*).

Taxonomic revision: Brandea. Bol Herb Bradeanu. 1991;5:1-20.

- Soares FP, Cavalcante LF, Romero NR, Bandeira MAM. Himatanthus Willd ex Schult. (Apocynaceae). Pharmacogn Rev. 2016;10(19):6. Available:https://doi.org/10.4103/0973-7847.176549
- Spina AP, Bittrich V, Kinoshita LS. Estudos taxonômico, micro-morfológico e filogenético do gênero Himatanthus Willd. ex Schult.(*Apocynaceae*: Rauvolfioideae-Plumerieae); 2004. Available:http://repositorio.unicamp.br/han dle/REPOSIP/315288
- Morais FS, Canuto KM, Ribeiro PRV, Silva AB, Pessoa ODL, Freitas CDT, et al. Chemical profiling of secondary metabolites from *Himatanthus drasticus* (Mart.) Plumel latex with inhibitory action against the enzymes α-amylase and αglucosidase: *In vitro* and in silico assays. J Ethnopharmacol. 2020;253:112644. Available:https://doi.org/10.1016/j.jep.2020 .112644
- 27. Santos GJL, Oliveira ES, Pinheiro ADN, Costa PM, Freitas JCC, Santos FGA, et al. *Himatanthus drasticus (Apocynaceae)* látex reduces oxidative stress and modulates  $CD_4^+$ ,  $CD_8^+$ ,  $FoxP_3^+$  and HSP- $60^+$  expressions in Sarcoma 180-bearing mice. J Ethnopharmacol. 2018;220:159-168.

Available:https://doi.org/10.1016/j.jep.2017 .09.043

- Moura DF, Rocha TA, Barros DM, Silva MM, Lira MAC, Souza TGS, et al. Evaluation of the cytotoxicity, oral toxicity, genotoxicity, and mutagenicity of the latex extracted from *Himatanthus drasticus* (Mart.) Plumel. J Ethnopharmacol. 2020; 253:112567. Available:https://doi.org/10.1016/j.jep.2020 .112567
- 29. Lucetti DL, Lucetti EC, Bandeira MAM, Veras HN, Silva AH, Leal LKA, et al. Antiinflammatory effects and possible mechanism of action of lupeol acetate isolated from *Himatanthus drasticus* (Mart.) Plumel. J Inflamm. 2010;7:60. Available:https://doi.org/10.1186/1476-9255-7-60
- Mousinho KC, Oliveira CC, Ferreira JRO, Carvalho AA, Magalhães HIF, Bezerra DP, et al. Antitumor effect of laticifer proteins of *Himatanthus drasticus* (Mart.) Plumel –

Apocynaceae. J Ethnopharmacol. 2011; 137(1):421-426.

Available:https://doi.org/10.1016/j.jep.2011 .04.073

- Almeida SCX, Monteiro, AB, Costa, GM, Viana GSB. *Himatanthus drasticus*: A chemical and pharmacological review of this medicinal species, commonly found in the Brazilian Northeastern region. Rev Bras Farmacogn. 2017;27(6):788-793.. Available:https://doi.org/10.1016/j.bjp.2017 .10.002
- Figueiredo CSSES, Santos JCB, Castro Junior JAA, Wakui VG, Rodrigues JFS, Arruda MO, et al. *Himatanthus drasticus* leaves: Chemical characterization and evaluation of their antimicrobial, antibiofilm, antiproliferative activities. Molecules. 2017;22(6)910. Available:https://doi.org/10.3390/molecules 22060910
- 33. Rebouças SO, Silva J, Bertoni RS, Decker N, Santos MS, Rossatto RR, et al. Assessment of the genotoxic and mutagenic properties of *Himatanthus articulatus* bark extracts used as phytotherapeutic drug in the Amazon. J Ethnopharmacol. 2013;147(2):474-480. Available:https://doi.org/10.1016/j.jep.2013 .03.041
- 34. Serqueira BJ, Vital MJS, Pohlit AM, Pararols IC, Caúper GSB. Antibacterial and antifungal activity of extracts and exudates of the Amazonian medicinal tree *Himatanthus articulares* (Vahl) Woodson (common name: sucuba). Mem Inst Oswaldo Cruz. 2009;104(4):659-661. Available:https://doi.org/10.1590/s0074-02762009000400022
- Rebouças SO, Grivicich I, Santos MS, Rodriguez P, Gomes MD, Oliveira SQ, et al. Antiproliferative effect of a traditional remedy, *Himatanthus articulatus* bark, on human cancer cell lines. J Ethnopharmacol. 2011;137(1):926-929. Available:https://doi.org/10.1016/j.jep.2011 .06.006
- Rattmann YD, Terluk MR, Souza WM, 36. Santos CAM, Biavatti MW, Torres LB, et al. Effects of alkaloids of Himatanthus lancifolius (Muell. Arg.) Woodson. on smooth Apocynaceae. muscle responsiveness. J Ethnopharmacol. 2005;100:268-275. Available:https://doi.org/10.1016/j.jep.2005 .02.041

- Baggio CH, Otofuji GM, Souza WM, Santos CAM, Torres LMB, Rieck L, et al. Gastroprotective mechanisms of indole alkaloids from *Himatanthus lancifolius*. Planta Med. 2005;71(8):733-738. Available:https://doi.org/10.1055/s-2005-871286
- Seidl C, Correia BL, Stinghen AEM, Santos CAM. Acetylcholinesterase inhibitory activity of uleine from *Himatanthus lanciflolius*. Z Naturforsch C J Biosci. 2010;65(7-8):440-444. Available:https://doi.org/10.1515/znc-2010-7-804
- Nardin JM, Souza WM, Lopes JF, Florão A, Santos CAM, Weffort-Santos AM. Effects of *Himatanthus lancifolius* on human leukocyte chemotaxis and their adhesion to integrins. Planta Med. 2008;74(10):1253-1258. Available:https://doi.org/10.1055/s-2008-1074582
- Souza WM, Stringhen AE, Santos CA. Antimicrobial activity of alkaloidal fraction from barks of *Himatanthus lancifolius*. Fitoterapia. 2004;75(7-8):750-753. Available:https://doi.org/10.1016/j.fitote.20 04.09.012
- 41. Silva ML, Stehamann JR, Serafim MSM, Vale VV, Gontijo DC, Brandão GC, et al. *Himatanthus bracteatus* stem extracts present anti-flavivirus activity while an isolated sesquiterpene glucoside present only anti-Zika virus activity in vitro. Nat Prod Res. 2019;20:1-5. Available:https://doi.org/10.1080/14786419 .2019.1690487
- Lorenzi H, Matos FJ. Plantas medicinais no Brasil: Nativas e Exóticas; 2002. Cronquist A. The evolution and classification of flowering plants. 2<sup>nd</sup> ed. New York: The New York Botanical Garden; 1988.
- Ferreira C, Piedade MTF, Parolin P, Barbosa KM. Tolerância de *Himatanthus sucuuba* Wood. (Apocynaceae) ao alagamento na Amazônia Central. Acta Bot Brasilica. 2005;19(3):425-429.

Available:https://doi.org/10.1590/S0102-33062005000300002

 Van Den Berg ME. Plantas medicinais na Amazônia: contribuição ao seu conhecimento sistemático. In: Plantas medicinais na Amazônia: contribuição ao seu conhecimento sistemático. 1982;223-223.

- Rios MNS, Pastore Júnior F. Plantas da Amazônia: 450 espécies de uso geral; 2011. Available:https://repositorio.unb.br/handle/ 10482/35458
- Lorenzi H. Arvores brasileiras manual de identificação e cultivo de plantas arbóreas do Brasil; 1998.
- 47. Larrosa CRR, Duarte M. Morfoanatomia de folhas de *Himatanthus sucuuba* (Spruce) Woodson, Apocynaceae. Acta Farmaceutica Bonaerense. 2005;24(2): 165. Available:https://docs.ufpr.br/~marcia/artig

Available:https://docs.ufpr.br/~marcia/artig os/folsucu.pdf

- 48. Di Stasi LC, Hiruma-Lima CA. Plantas medicinais na Amazônia e na Mata Atlântica. Editora Unesp; 2002.
- Miranda ALP, Silva JRA, Rezende CM, Neves JS, Parrini SC, Pinheiro MLB, et al. Anti-inflammatory and analgesic activities of the latex containing triterpenes from *Himatanthus sucuuba*. Planta Med. 2000; 66(3):284-286. Available:https://doi.org/10.1055/s-2000-

8572

 Soares DC, Andrade ALS, Delorenzi JC, Silva JRA, Freire-De-Lima L, Falcão CAB, et al. Leishmanicidal activity of *Himatanthus sucuuba* latex against Leishmania amazonensis. Parasitol Int. 2010;59(2):173-177. Available:https://doi.org/10.1016/i.parint.20

Available:https://doi.org/10.1016/j.parint.20 10.01.002

- 51. Rapa SF, Waltenberger B, Paola RD, Adesso S, Siracusa R, Peritore AF, et al. Plumericin prevents intestinal inflammation and oxidative stress *In vitro* and in vivo. The FASEB Journal. 2020;34(1): 1576-1590. Available:https://doi.org/10.1096/fj.201902 040R
- 52. Souza KP. Propagação *in vitro* de *Himatanthus sucuuba* Wood, uma espécie medicinal da Amazônia. (PhD in Biotechnology and Natural Resources), Universidade do Estado do Amazonas, Manaus; 2017. Available:http://www.pos.uea.edu.br/data/a rea/titulado/download/71-7.pdf
- 53. Soares TS, Damasceno DC, Kempinas WG, Resende FMC, Santos MAC, Hiruma-CA. et al. Fffect of Lima Himatanthus sucuuba in maternal reproductive outcome and fetal anomaly frequency in rats. Birth Defects Res (Part B). 2015;104:190-195.

Available:http://dx.doi.org/10.1002/bdrb.21 152

- Villegas LF, Fernandez ID, Maldonado H, Torres R, Zavaleta A, Vaisberg AJ, et al. Evalution of the wound-healing activity of selected traditional medicinal plants from Peru. J Ethnopharmacol. 1997;55:193-200. Available:https://doi.org/10.1016/s0378-8741(96)01500-0
- Wood CA, Lee K, Vaisberg AJ, Kingston DG, Neto CC, Hammond GB. A bioactive spirolactone iridoid and triterpenoids from *Himatanthus sucuuba*. Chem Pharm Bull. 2001;49:1477-1478.

Available:https://doi.org/10.1248/cpb.49.14 77

- 56. Castillo D, Arevalo J, Herrera F, Ruiz C, Rojas R, Rengifo E, et al. Spirolactone iridoids mignt be responsible for the antileishmanial activity of a Peruvian traditional remedly made with *Himatanthus sucuuba* (Apocynaceae). J Ethnopharmacol. 2007;112:410-414. Available:https://doi.org/10.1016/j.jep.2007 .03.025
- Rodrigues E, Duarte-Almeida JM, Pires JM. Phytochemical and pharmacological profile of plants indicated by caboclos of Jaú National Park (AM) as potential analgesic. Part I. Rev Bras Farmacogn. 2010;20(6):981-991. Available:https://doi.org/10.1590/S0102-695X2010005000008
- 58. Pires LC, Pereira MFS, Martins ST, Tómaz LAD, Ferreira FAG. Etnologia e atividade biológica da planta medicinal *Himatanthus sucuuba*. Scientia Prima. 2016;4(4):5-10. Available:https://issuu.com/scientiaprima/d ocs/scientia prima vol4 n4
- 59. Paz MFCJ, Alencar MVOB, Soares RLL, Costa DAF, Nunes AT, Cavalcante AACM. Evaluation toxic, cytotoxic, mutagenic and genotoxic látex *Himatanthus sucuuba*: A public health issue. Revista Interdisciplinar. 2013;6(1):52-61. Available:https://revistainterdisciplinar.unin

ovafapi.edu.br/index.php/revinter/article/vie w/10/pdf\_4

- Neto CC, Owens CW, Langfield RD, Comeau AB, Onge JS, Vaisber AJ, et al. Antibacterial activity of some medicinal plants from the Callejon de Huaylas. J Ethnopharmacol. 2002;79:133-138. Available:https://doi.org/10.1016/S0378-8741(01)00398-1
- 61. Endo Y, Hayashi H, Sato T, Maruno M, Ohta T, Nozoe S. Confluentic Acid and 2'-

O-Methylperlatolic acid, monoamine oxidase B inhibitors in a Brazilian plant, *Himatanthus sucuuba*. Chem Pharm Bull. 1994;42(6):1198-1201. Available:https://doi.org/10.1248/cpb.42.11 98

 Silva JRA, Rezende CM, Pinto AC, Pinheiro MLB, Cordeiro MC, TamboriniE, YoungCM, BolzaniVS. Ésteres triterpênicos de *Himatanthus sucuuba* (Spruce) Woodson. Quim Nova. 1998;21: 702-704. Available:https://doi.org/ 10.1590/S0100-

40421998000600005 Souza MS, Cordeiro MS, Rosas EC, Henriques MGOM, Saini AC. Inhibition of

63.

Herinques MGOM, Sam AC. Inhibition of nitric oxide and interferon-γ production by iridoids and triterpenes from the roots of *Himatanthus sucuuba*. Phcog Mag. 2006; 2(8):216-219.
 Available:https://citeseerx.ist.psu.edu/view

doc/download?doi=10.1.1.115.7772&rep=r ep1&type=pdf

64. Fakrudin N, Waltenberger B, Cabaravdic M, Atanasov AG, Malainer C, Schachner D, et al. Identification of plumericin as a potent new inhibitor of the NF-kB pathway with anti-inflammatory activity in vitro and in vivo. Br J Pharmacol. 2014;171(7):1676-1686.

Available:https://doi.org/10.1111/bph.1255 8

- Morel AF, Graebner IB, Porto C, Dalcol II. Study on the antimicrobial activity of *Hymatanthus sucuba*. Fitoterapia. 2006;77:50-53. Available:https://doi.org/10.1016/j.fitote.20 05.08.012
   Silva JRA, Rezende CM, Pinto AC, Amaral
- Silva JRA, Rezende CM, Pinto AC, Amaral ACF. Cytotoxicity and antibacterial studies of iridoids and phenolic compounds isolated from the latex of *Himatanthus sacuuba*. Afr J Biotechnol. 2010;9:7357-7360.

Available:https://doi.org/10.5897/AJB10.34 5

- 67. Kumar R. Reeta KH. Rav SB. Antinociceptive effect of intrathecal loperamide: role of mu-opioid receptor and calcium channels. Eur J Pharmacol. 2012;696:77-82. Available:https://doi.org/10.1016/j.ejphar.2 012.09.022
- Norton RS, Pallaghy PK, Baell JB, Wright CE, Lew MJ, Angus JA. Polypeptide ω-conotoxin GVIA as a basis for new

analgesic and neuroprotective agents. Drug Dev Res. 1999;46(3-4):206-218. Available:https://doi.org/10.1002/(SICI)109 8-2299(199903/04)46:3/4<206::AID-DDR6>3.0.CO:2-4

- Florentino IF, Nascimento MVM, Galdino PM, Brito AF, Rocha FF, Tonussi CR, et al. Evaluation of analgesic and antiinflammatory activities of *Hydrocotyle umbellate* L., Araliaceae (acariçoba) in mice. An Braz Acad Scienc. 2013;85(3): 987-997. Available:https://doi.org/10.1590/S0001-37652013000300011
- 70. Bhowmick R, Sarwa MS, Dewan SMR, Das B, Uddin MMN, Islam MS, et al. *In* vivo analgesic, antipyretic, and antiinflammatory potential in Swiss albino mice and *in vitro* thrombolytic activity of hydroalcoholic extract from *Listea glutinosa* leaves. Biol Res. 2014;47:56. Available:http://dx.doi.org/10.1186/0717-6287-47-56
- Azmi F, Skwarczynski M, Toth I. Towards the development of synthetic antibiotics: Designs inspired by natural antimicrobial peptides. Curr Med Chem. 2016;23(41): 4610-4624. Available:https://doi.org/10.2174/09298673 23666160825162435
- Nascimento GGF, Locatelli J, Freitas PC, Silva GL. Antibacterial activity of plant extracts and phytochemicals on antibioticresistant bacteria. 2000;31(4):247-256. Available:http://dx.doi.org/10.1590/S1517-8382200000400003
- 73. Mostafa AA, Al-Askar AA, Almaary KS, Dawoud TM, Sholkamy EN, Bakri MM. Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. Saudi J Biol Sci. 2018; 25(2):361-366. Available:https://doi.org/10.1016/j.sjbs.201
- 7.02.004
  74. Yamaguchi KKL, Souza AO. Antioxidant, hypoglycemic and neuroprotective activities of extracts from fruits native to the Amazon region: A review. Biotechnol J Int. 2020;24(6):9-31. Available:https://doi.org/10.9734/BJI/2020/ v24i630119
- 75. Hwang JS, Lee SA, Hong SS, Lee KS, Lee MK, Hwang BY, et al. Monoamine oxidase inhibitory components from the roots of Sophora flavescens. Arch Pharm Res. 2005;28(2):190-194.

Available:https://doi.org/10.1007/BF02977 714

- Herraiz T, Guillén H. Monoamine oxidase-A inhibition and associated antioxidant activity in plant extracts with potential antidepressant actions. BioMed Res Int. 2018;4810394:10. Available:https://doi.org/10.1155/2018/481 0394
- 77. Mazu TK, Bricker BA, Flores-Rozas H, Ablordeppey SY. The mechanistic targets of antifungal agents: An overview. Mini Rev Med Chem. 2016;16(7):55-578. Available:https://doi.org/10.2174/13895575 16666160118112103
- Kumarasingha R, Preston S, Yeo TC, Lim DSL, Tu CL, Palombo EA, et al. Anthelmintic activity of selected ethnomedicinal plant extracts on parasitic stages of *Haemonchus* contortus. Parasites Vectors. 2016;9:187. Available:https://doi.org/10.1186/s13071-016-1458-9
- Almeida Júnior S. *In vivo* methods for the evaluation of anti-inflammatory and antinoceptive potential. BrJP. 2019;2(4): 386-389. Avaliable:https://doi.org/10.5935/2595-0118.20190070
- Zeeli S, Weill T, Finkin-Groner E, Bejar C, Melamed M, Furman S, et al. Synthesis and biological evaluation of derivates of indoline as highly potent antioxidant and anti-inflammatory agents. J Med Chem. 2018;61(9):4004-4019. Available:https://doi.org/10.1021/acs.jmedc hem.8b00001
- Kure C, Timmer J, Stough C. The immunomodulatory effects of plant extracts and plant secondary metabolites on chronic neuroinflammation and cognitive aging: A mechanistic and empirical review. Front Pharmacol. 2017;8:117. Available:https://doi.org/10.3389/fphar.201 7.00117
- Anversa L, Tiburcio MGS, Richini-Pereira VB, Ramirez LE. Human leishmaniasis in Brazil: A general review. Rev Assoc Med Bras. 2018;64(3):281-289. Available:http://dx.doi.org/10.1590/1806-9282.64.03.281
- 83. Santos AL, Yamamoto ES, Passero LFD, Laurenti MD, Martins LF, Lima ML, et al. Antileishmaninal activity and immunomodulatory effects of tricin isolated from leaves of *Casearia arborea*

(Salicaceae). Chem Biodivers. 2017;14(5): e1600458.

Available:https://doi.org/10.1002/cbdv.201 600458

 Bhalla G, Kaur S, Kaur J, Kaur R, Raina P. Antileishmanial and immunomodulatory potential of *Ocimum sanctum* Linn. and *Cocos nucifera* Linn. In murine visceral leishmaniasis. J Parasit Dis. 2017;41(1): 76-85.

Available:https://doi.org/10.1007/s12639-016-0753-x

85. Dey S, Mukherjee D, Chakraborty S, Mallick S, Dutta A, Ghosh J, et al. Protective effect of *Croton caudatus* Geisel leaf extract against experimental visceral leishmaniasis induces proinflammatory cytokines *in vitro* and *in vivo*. Exp Parasitol. 2015;151-152:84-95. Available:https://doi.org/10.1016/j.

exppara.2015.01.012

 Teng YN, Wang Y, Hsu PL, Xin G, Zhang Y, Morris-Natschke SL, et al. Mechanism of action of cytotoxic compounds from the seeds of *Euphorbia lathyris*. Phytomedicine. 2018;41:62-66. Available:https://doi.org/10.1016/j.phymed. 2018.02.001

- Çevik D, Kan Y, Kirmizibekmez H. Mechanisms of action of cytotoxic phenolic compounds from *Glycyrrhiza iconica* roots. Phytomedicine. 2019;58:152872. Available:https://doi.org/10.1016/j.phymed. 2019.152872
- Mazzu-Nascimento T, Melo DG, Morbioli GG, Carrilho E, Vianna FSL, Silva AA, et al. Teratogens: A public health issue – a Brazilian overview. Genet Mol Biol. 2017;40(2):387-397. Available:http://dx.doi.org/10.1590/1678-4685-GMB-2016-0179
- Araújo CRF, Santiago FG, Peixoto MI, Oliveira JOD, Coutinho MS. Use of medicinal plants with teratogenic and abortive effects by pregnant women in a city in Northeastern Brazil. Rev Bras Ginecol Obstet. 2016;38:127-131. Available:http://dx.doi.org/ 10.1055/s-0036-1580714.

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