



A Comprehensive Review on Melatonin Compound and its Functions in Different Fungi and Plants

Rao Saad Rehman ^{a*}, Mubashar Hussain ^a, Mujahid Ali ^b, Syed Ali Zafar ^c,
Asad Nadeem Pasha ^d, Hassan Bashir ^e, Naveed Ali Ashraf ^f,
Abdullah Javed ^f and Waqar Ali Shah ^f

^a College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China.

^b Department of Plant Breeding and Genetics, Nanjing Agricultural University, Nanjing, China.

^c Oilseeds Research Institute, Ayub Agricultural Research Institute, Faisalabad, Pakistan.

^d Department of Plant Pathology, Bahauddin Zakariya University, Multan, Pakistan.

^e Department of Plant Breeding and Genetics, Sub Campus Burewala-Vehari,
University of Agriculture Faisalabad, Pakistan.

^f Department of Seed Science and Technology, University of Agriculture Faisalabad, Pakistan.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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Review Article

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ABSTRACT

This study summarizes the importance of melatonin in different plants and fungi. In this review, we discussed the biosynthetic pathway of melatonin, its metabolites, and its oxidative reduction. Melatonin is a molecule derived from tryptophan, with pleiotropic activity. It is present in nearly every organism. Its synthetic course depends on the organism in which it resides. The tryptophan to the melatonin pathway, for example, varies in plants and animals. It is thought that the synthetic mechanism for melatonin was inherited in eukaryotes from bacteria caused by endosymbiosis. Nevertheless, the synthetic pathways of melatonin in microorganisms are unknown. The metabolism of melatonin is exceptionally complex with these enzymatic processes developed out of cytochrome C. As well as the enzymatic degradation, melatonin is metabolized by interactive pseudoenzymes and free radicals processes.

Keywords: *Melatonin; biosynthetic pathway; photoperiodic changes; suprachiasmatic nucleus; reactive oxygen species; heat tolerance.*

1. INTRODUCTION

Melatonin is a small compound that was first identified in the pineal glands of cows [1]. Its core seen task was to make amphibians skin lighter by causing granules of melanin compound granules inside dermal melanosomes to be aggregated in frogs around the nucleus of skin cells [2]. The identified molecule was therefore called melatonin because it forces dark pigment (melanin) to lighten in the skin and is often extracted out from serotonin. Due to that action, in certain patients with concentrated hyperpigmented skin, melatonin was once used to minimize pigmentation. This was found to be not successful without a skin lightening effect in human beings [3]. The point is that mammalian melanosomes are more or less like those found in amphibians dispersed indefinitely, and therefore, melatonin has no impact on its capacity to change pigment aggregation in mammalian Skin. Some of the most unusual characteristics of melatonin in vertebrates are its circadian cycle, including its secretory night peak, and low daytime levels. It makes melatonin an effective signaling agent to inform internal organs of photoperiodic changes to the environment. At the end of melatonin in vertebrate blood, melatonin often correlates with the dark period of the light/night cycle, referred to as dim chemical expression [4]. The normal circadian melatonin rhythm in vertebrates is created solely by melatonin excreted from the pineal gland, after either pineal ectomy vanishes this pattern in the blood [5], or growing its intensity varies significantly [6]. The master clock, the pineal gland, and the suprachiasmatic nucleus (SCN) interact with one another, consist of key bio-clock elements that synchronize the physiological activities of organisms, most importantly in vertebrates with predominant light/dark cycle [7, 8]. SCN relays details concerning photoperiods using the sympathetic nervous system to the pineal gland and relies on the details received by the SCN, the pineal gland regulates the production of melatonin either up- or down-. Because the melatonin function is changing. When it is discharged, it affects the functioning of SCN [9, 10]. Receptors of melatonin, most importantly MT1, are appeared in SCN. Melatonin receptors, especially MT1, are expressed in the SCN [11, 12] and are densely populated. The interactions have been between SCN, sympathetic nervous system, pineal gland

[13–15]. Changes in the levels of melatonin due to diurnal are apparent in photosynthetic primitive bacteria [16, 17] in which they are identical to those found in vertebrate species; but, these diurnal changes can occur be passive, that is, the low melatonin levels in bacteria during photo phase may not be the result from a reduced synthesis of this indole but also oxidation as a consequence of its application. It is being hypothesized that more melatonin in photosynthetic organisms is metabolized during the day because of its reactive oxygen species interaction (ROS). This is because bacteria are photosynthetic, photosynthesis is maximum during the day, and produces large amounts of ROS through this process. Moreover, the diurnal modification of melatonin in prokaryotes does not have a direct effect on the role of their bio-clock, but the shifts are simply a result of their irregular metabolism that happens before and throughout the day [18]. We had a clear one diel melatonin rhythm in a single-celled organism, *Symbiodinium* genus dinoflagellate. Throughout this sense species, the peak of melatonin occurred in the dark with low daytime levels. These changes are disappeared when the organism is placed in the dark. Based on modifications to the production of oxygen during the light/dark cycle resulting from photosynthesis in this organism, the researchers finally concluded that fluctuating pattern in *symbiodinium* was not induced by endogenous circadian synthesis, however, also shifts in its application owing to the regular photocycle. The rhythm was attributable to pathways involving increased light-utilization of melatonin through the use of free radicals in the morning. The evident circadian melatonin rhythm of vertebrates probably evolved during evolution from the passive improvements in the development of melatonin in bacteria; the rhythm then evolved to influence the bio-clock of Superior organisms [19]. For example, circadian melatonin rhythm relating to the sleep cycle is *platynereis dumerilii*, already observed in early marine zooplankton [20]. It could be that organism, the first animal in which melatonin acted as a quiescent signal molecule. Disorders in the circadian cycle of melatonin result in chronodisruption that is consistent with many health conditions, including neurodegenerative diseases, heart disease, high blood pressure, and cancer. For example, shift workers, notably nurses, aircraft crews, and miners, have a higher breast and prostate cancer

prevalence [21–23]. One possible mechanism is light exposure during the night (their operating hours) suppresses the melatonin levels [24]. Which is compatible with the statements in the animal in which human donors infuse tumor-bearing rats with melatonin-rich blood the development of the transplanted tumor (having nightly melatonin levels) was inhibited; however, if these rats were injected with equivalent daytime human blood deficient in melatonin, tumor development was released [25]. Therefore, as light exposure exaggerates tumor growth during the dark process. Melatonin production also exhibits seasonal changes as a signaling molecule. In the winter, the late peak period of melatonin is longer due to the longer nights, during the summer, the night-time high is shorter [26, 27]. Photoperiodic animals use this information to make adjustments to their reproductive behaviors during the correct season [28] and their hibernation behavior [29]. For example, without a proper melatonin message after pinealectomy, which through the blood melatonin rhythm, reproductive disturbances, and the hibernating photoperiodic cycle is noticeable [29–31]. The circadian rhythm of melatonin is neither as clear nor regular in plants as it is in animals. Only increased amounts of melatonin were observed during scotophase, and lower amounts during photophase *Chenopodium rubrum*, in an evolutionarily ancient plant [32]. Indeed, multiple reports have shown that light-exposure induces the development of melatonin in plants [33–35]. If there is more sunlight, then there is more synthesis of melatonin molecules in

a few plants. Photosynthesis is believed to produce significant amounts of ROS and plants; more melatonin is probably developed to defend against toxicity [36, 37]. Free radical scavenger and antioxidant are core functions of melatonin. While other melatonin functions have been acquired in all organisms in development [19, 38]. Because melatonin is among those products which are found naturally, so it is different in many ways from commonly found antioxidants. These involve melatonin cascade reaction with ROS and the possibility that its synthesis is inducible to high oxidizing stress in humans. These unique characteristics of melatonin make it bigger suitable for the defense of species from stressful conditions as an endogenous antioxidant.

2. BIOSYNTHETIC PATHWAY OF MELATONIN

The Source of melatonin in animals is the pineal gland. However, that organ does not exist in plants, and this discrepancy suggests that cell melatonin biosynthesis is somewhat different than in animals [39]. In plants, melatonin is found in different parts of plants like root, stem, and leaf, unlike in the animals. In plants, multiple factors may stimulate the biosynthesis of melatonin. Light is among the environmental factors which regulate melatonin biosynthesis [40]. There are more growth processes such as fruit ripening [41], plant development [42], and senescence [43], and climate stresses involved

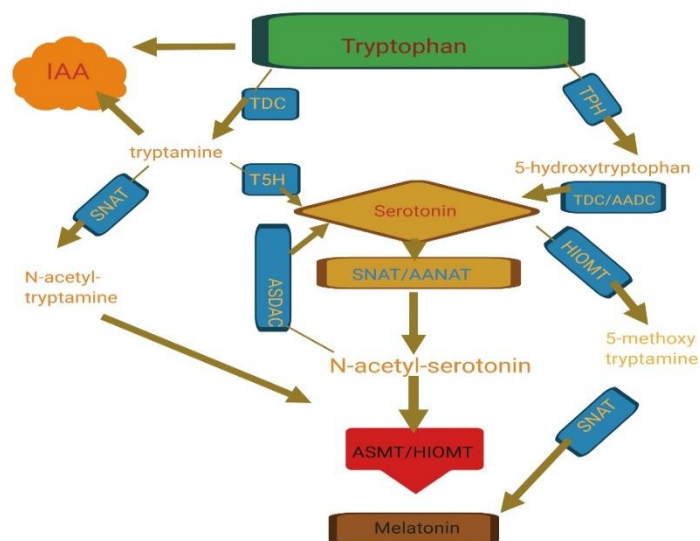


Fig. 1. The biosynthetic pathway of melatonin in plants

include Ultraviolet-B (UV-B) radiation [44], drought, cold [45] and heat [46] that stimulate melatonin biosynthesis. Melatonin biosynthesis begins in a wide array of plant species through tryptophan. Tryptophan decarboxylase (TDC) is catalyzed and transformed to tryptamine, as shown in Fig. 1; next, tryptamine 5-hydroxylase (T5H) catalyzes tryptamine in serotonin, which is transformed by two steps to melatonin [47]. Among certain other species, tryptophan, such as *Hypericum perforatum*, is catalyzed by tryptophan 5-hydroxylase (TPH) in 5-hydroxytryptophan, and then TDC / AADC (Decarboxylase, Aromatic-L-amino acid) transforms 5-hydroxytryptophan to serotonin [48]. This path is the same true of animals. The serotonin is converted to N-acetyl-serotonin in the next two steps using serotonin N-acetyltransferase (SNAT)/arylalkylamine N-acetyltransferase (AANAT), and then catalysis of n-acetyl-serotonin methyltransferase (ASMT)/hydroxyindole-O-methyltransferase (HIOMT) N-acetyl-serotonin comes in melatonin. Additionally, SNAT can be catalyzed from tryptamine into N-acetyl-tryptamine, which T5H [49] does not further convert into N-acetyl-serotonin. This is difficult to determine whether a pathway for converting N-acetyl-tryptamine to N-acetyl-serotonin exists. The other route is by HIOMT to transform serotonin into 5-methoxy-tryptamine and eventually catalysis of 5-methoxy-tryptamine by SNAT into melatonin [50, 51]. A reverse melatonin pathway was recently discovered. Reported, in which N-acetyl-serotonin is converted by N-acetyl-serotonin deacetylase into serotonin [52]. Tryptophan is not only a provider of melatonin but also an indole-3-acetic precursor acid (IAA), perhaps implying melatonin multifunctional role in plants.

3. METABOLISM OF MELATONIN

Melatonin oxidation is less well understood than melatonin synthesis. For several decades, the only significant metabolite was 6-hydroxymelatonin and, therefore, several studies centered on it. Melatonin synthesis is a full cycle, and 6-hydroxymelatonin is one of its metabolites. Melatonin metabolizes through enzymatic mechanisms, pseudo enzymatic mechanisms, or encounters with ROS and NOS. Melatonin is found in primary photosynthetic bacteria like the prokaryote *rhodospirillum rubrum*, cyanobacteria [53]. The former is the parent to mitochondria, occurring in nearly all cells, is the possible source to chloroplasts seen in green plants. There is not that much research on the

metabolism of melatonin. Nonetheless, melatonin synthesis in mitochondria is established. In mitochondria, melatonin is degraded through the pseudo-enzymatic method. Cytochrome C acts as an enzyme to eliminate melatonin in N1-acetyl-N2-formyl-5-methoxykynuramine.

Cytochrome C potent independent protein, even in bacteria [54]. Therefore, we believe the bacteria will even use melatonin-metabolizing cytochrome C. Cytochrome C may be the first degrading protein degrading melatonin, and AFMK may be the first pseudo enzymatic result of the melatonin metabolism process. Method. The cytochrome C catalytic core for melatonin metabolism can be in its molecule of a single iron atom.

Such iron-containing hemoproteins such as AFMK and other metabolites associated with melatonin [55]. In this cycle, melatonin is oxidized and cleaned to form AFMK by oxoferryl-hemoprotein [56]. Many enzymatic pathways involved in melatonin synthesis during development. Most enzymes responsible for melatonin synthesis have a specific feature: iron-containing cytochrome-like hemoproteins. Namely cytochrome P450 (CP450), indole amine 2,3-IDO, horseradish peroxidase (HRP), myeloperoxidase (MPO), and eosinophilic peroxidase. Every enzyme will break melatonin into AFMK. CP450 is the animal's main enzyme for melatonin synthesis. It is primarily in the liver, but often present in many tissues. CP450 primarily generates 6-hydroxymelatonin, and its small product is AFMK. In the brain, the primary enzyme for melatonin synthesis is ID with its product AFMK. MPO and EPO are responsible for melatonin synthesis at inflammatory sites. HRP exists in plants and can engage in plant melatonin metabolism.

AFMK line, plants identified. Cyclic melatonin (unlike cyclic-3-hydroxymelatonin) and β -hydroxy melatonin were detected in plants as shown in Fig. 2.

Dominant melatonin in plants (24 species) metabolite is 2-hydroxymelatonin and is known as melatonin-2-hydroxylases [57]. When melatonin is made, it rapidly becomes 2-hydroxymelatonin. In the plants examined, the normal ratio of melatonin to 2-hydroxymelatonin was 1:368. If the ratio is verified according to others, melatonin synthetic ability in plants is much more effective than past aspirations. Interestingly, a non-enzymatic mechanism may also metabolize melatonin. Melatonin complex

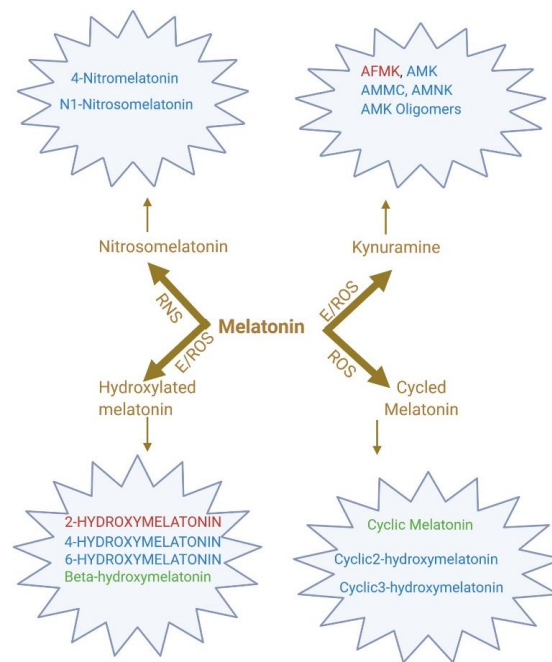


Fig. 2. Metabolites of melatonin; blue metabolites are found only in animals, green are found in plants and red are found in both animals and plants

interactions with ROS and NOS produce different metabolites. These contain 3-hydroxymelatonin, 2-Hydroxymelatonin, N-nitrosomelatonin, N-(1-formyl-5-methoxy-3-oxo-2,3-Dihydro-1H-indol-2) acetamide, AFMK, and AMK [58]. Enzyme-related metabolites, pseudo enzymatic processes, and ROS interaction pathways are common.

Furthermore, it is complicated to determine through the metabolic cycle that dominates in vivo conditions. In extreme oxidation stress, however, it is believed that ROS interactions metabolize most melatonin because of oxidative stressors dramatically lower overall organism melatonin rates. The melatonin synthesis nature faces problems for researchers. None is known to date. There is almost little research about melatonin synthesis in plant microorganisms. We assume in melatonin synthesis and several other pathways melatonin metabolites should be identified in the coming years [65].

4. ANTIOXIDANT ACTIVITY OF MELATONIN

While melatonin is found to be a decade earlier, a free radical scavenger [59], the data that recorded its function to conquer stress due to oxidative has accrued at an accelerated rate. The rate is now abundant [60-62]. Efficacy of

melatonin for working throughout this power, it relates to its parent-free radical scavenging behavior, the capacity to improve the behaviors of several individuals. Antioxidant enzymes, it is a relaxing impact on the synthesis of another essential intracellular antioxidant, glutathione its effectiveness in the elimination of nucleus loss of the nuclear nucleus transport chain [63] and synergistic chain many antioxidant associations [64]. Moreover, it has been evident in recent years this is why melatonin scavenges radicals, related reactants, the products that are produced open radical scavengers, however, thereby massively exaggerating melatonin antioxidant potential [66]. Subsequent articles shall contain a summary of the melatonin-capable metabolites neutralizing free radicals and oxygen-dependent non-radicals the reagents [67].

4.1 Melatonin Reduces the Oxidative Stress Induced by Hydrogen Peroxide in *saccharomyces*

Melatonin (N-acetyl-5-methoxytryptamine) has a tryptophan synthesis cerevisiae and unusual yeast genus *Saccharomyces*. Properties of antioxidant the role of melatonin in an *S. cerevisiae* were suggested as possible wine strain. The possible effect on non-*Saccharomyces* species of antioxidants melatonin and other strains. It is necessary to

evaluate cerevisiae. The object of this analysis [68] was to determine the melatonin antioxidant capacity in 8 *S.* strains and cerevisiae four yeasts (*Torulaspora delbrueckii*, *Metschnikovia pulcherrima*, *Hanseniaspora uvarum*, and *Starmerella bacillaris*). The formation of ROS, therefore fat peroxidation, operation of catalase, fatty acids, and proliferation of peroxisomes, the inquiry has been completed. The tests indicated an improvement in melatonin peroxisome build-up, and the operation of the catalase decreases considerably. When rising cells oxidative stress induced by H₂O₂ was exposed to melatonin, all tested strains have observed lower ROS accumulation and lipid peroxidation [69]. The decreased development of catalysis, which was a consequence of oxidative stress, was less in melatonin presence. Also, MEL modulates cell presence composition of FA, increased oleic and palmitoleic acids, and increased UFA / SFA ratios previously associated with increased H₂O₂ tolerance. These discoveries prove that melatonin can act as an antioxidant in both *S. cerevisiae* and yeasts, which are non-Saccharomyces [70].

4.2 Growth of *Xanthomonas oryzae* PV. *oryzae* is Inhibited by Melatonin

Xanthomonas oryzae PV. *oryzae* (Xoo), one of the many severe and devastating diseases that exist in rice-growing regions around the world. Melatonin improves pathogenic tolerance by causing plant innate immunity, which has clear effect melatonin is little known on plant pathogenic bacteria. In the analysis [71], the immediate influence of melatonin on Xoo has been studied. Exogenous, 200 mg / mL melatonin, the Xoo proliferation was significantly impaired, and the mRNA production decreased by five genes active in separating cells [72]. This melatonin production also impaired motility; and Xoo Biofilm Formation. Melatonin was noteworthy for altering the length of Xoo cells. Provide more insight into the mechanisms that underlie this antibacterial activity. Also, we examined changes in global gene expression of Xoo strain PXO99 using RNA sequencing (RNA-Seq) for application of 200 mg / mL melatonin. Differential gene range (DGGs) related to catalytic activity and metal binding in response to the melatonin treatment, activity in Xoo cells was de-regulated [73]. Additionally, DEGs responsible for the metabolism of carbohydrates and amino acids were down-regulated too. Those results suggest melatonin inhibitory mechanism on Xoo proliferation; cell division can be controlled in

conjunction with a reduction of the production or operation of metabolism enzymes [74].

5. ROLE OF MELATONIN IN PLANTS

Melatonin production has already been seen in a variety of plants, and [75] were regularly checked. Given the vast number of exhibitions, the awareness remarkably low on bodily roles; this was primarily because scholars became less involved in plant physiology with also to include medical or therapeutic goods many advantageous acts are ascribed to indole amine origins. A more Chrono biological function was pursued or little to no avail [76]. *Chenopodium rubrum* is an excellent research organism that was defined as a nocturnal peak circadian rhythm [77], but a photoperiodic short day presence response [78] remains vague. Same like, no short day in certain Lemnaceans and *Kalanchoë* responses were found [79] *Tubiflora*. For some other plants, the rhythms recorded in tomatoes such as these have not been authenticated, but instead changes during ripening of fruits [80]. A pattern well marked the *Eichhornia crassipes*, mentioned in the water hyacinth [81]. In this species, at the end of the photo phase, the limit was achieved, and followed by the periodicity of melatonin by another solid metabolite pattern AFMK. All have identified an auxin-like growth stimulus in a lupine, dicot [82], also in monocots. Stimulating progress in coleoptiles of many poacemen was related to another auxin-like operation, suppression of root growth [83-91]. Whether or not these results are essential in physiology and whether melatonin it can include metabolites, it is an interesting problem to address in the future. Particular involvement of high levels of melatonin in fruits, and especially oily seeds possible function in process separation and maintenance, maybe of dormancy, coupled with antioxidant dry seed defense, in which the enzymatic pathways cannot act [92]. In *Eichhornia*, particularly the high levels of AFMK which reached a limit at the end was detected photophase, i.e., a period at which light-induced harm photosystems and secondary pathways to recover are the best shapes, and hence oxidizing [93-99]. This may mean in the process of moderate incremental degradation of melatonin free radicals process, singlet oxygen, late photocatalytic hemoproteins, peroxidases, and other chemicals. On the other side, specific melatonin-consuming reactions can be deemed to be contributions to photoprotection. None, the function of melatonin in photoprotection, was thought to be one of the

animal tissue types, such as organs high in porphyrins photo catalytically healthy, like rodents harderian gland, certain mollusks in photoreceptors and crustaceans, and even in numerous macro- and microalgae [100]. This can extend similarly to higher plants [83], as shown in a variety of comments: I light-dependent melatonin recycling, (ii) UV-induced increases, and (iii) significantly; higher amounts of melatonin contained in exposed plants about a significant degree of natural radiation, for example in the Alps, the Mediterranean, and subtropical areas, along with the same ecosystem elsewhere or, in [84-87] greenhouses.

5.1 Melatonin Induces Heat Tolerance in *Arabidopsis*

Melatonin (N-acetyl-5-methoxytryptamine) is an essential part of the signal molecule in the process of plant development and multiple abiotic stress reactions. Melatonin involvement in thermos tolerance, however, in *Arabidopsis*, the fundamental molecular process was mainly not known. Shi [88] reported that the endogenous level of melatonin coupled with heat stress treatment induced *Arabidopsis* leaves significantly, and exogenous melatonin treatment enhanced thermos tolerance in *Arabidopsis*, which is [89]. The class A1 heat-shock factor transcript levels (HSFA1s), which act as master heat stress response regulators, have been essential, and in *Arabidopsis*, heat stress and exogenous melatonin treatment are upregulated. Of particular note, exogenous thermos tolerance enhanced by melatonin was enhanced mainly in HSFA1 quadruple knockout (QK) mutants, and HSFA1 activated heat-responsive gene transcripts (HSFA2), heat-induced stress response can be 32 (HSA32), 90 (HSP90) and 101 (HSP101) heat-shock protein has led to thermos tolerance regulated by melatonin [90, 91]. Some research provided a direct connection between melatonin and heat tolerance and demonstrated the engagement of heat-response genes activated by HSFA1s in thermos tolerance mediated with melatonin in *Arabidopsis* [101-103].

5.2 Melatonin Induces Heat Tolerance in Tomato Plants

Melatonin is a pleiotropic molecule for physiological protection compared to various environmental stresses in plants. The mechanisms for melatonin driven

thermotolerance are uncertain. [104] reported the number of endogenous melatonin increases as air temperature rises and that about 40 ° C. Optimal melatonin dose (10 µmol / L) foliar pretreatment or N-acetylserotonin over-expression effectively, DNA methyltransferase (ASMT) heat-induced change tomato photography and electrolyte leakage trees. Trees, [105]. Treatment of exogenous melatonin and manipulation of melatonin the levels of insoluble and ubiquitous proteins decreased by over-expression of ASMT, but improved heat-shock expression HSPs for refolding denatured proteins and heat-stressed unfolded proteins [106]. Melatonin has also led to expression. Meanwhile, multiple ATG genes and autophagosome formation aggregated to degrade under the same stress, proteins. Analysis of the proteomic profile found that protein aggregates accumulated in the wild for a large number of biological processes trees. Trees, [107]. The treatment or overexpression of ASMT in exogenous melatonin, however, was reduced aggregated protein accumulation. Protein reactors such as aggregation preference were given to the accumulation and ubiquitisation of HSP70 and Rubisco activase type Wild heat stress plants, while melatonin reduces heat stress aggregated protein accumulation and ubiquitisation [108]. These findings show that the combination of HSPs and melatonin facilitates cellular protein defense and autophagy in tomato plants for refolding or degrading heat stress denatured proteins [109, 110].

5.3 Melatonin Induces Heat Tolerance in Maize Seedlings

Melatonin (MT) is a superb signaling molecule with multiple functions in plants and derived from tryptophan. Induced thermal stress (HS) a significant stress factor limiting plant metabolism, growth, development, and productivity is high temperature. MT could improve maize seedlings thermotolerance, and the underlying mechanisms are unknown. In research [111] reported, treatment of maize seedlings with MTE the survival percentage of maize seedlings under HS has been improved and the conditions mitigated malondialdehyde and electric leakage and improved tissue vitality in comparison with MDA (membrane lipid peroxidation) treatment control without MT indicates that the thermotolerance of maize seedlings could be enhanced by MT treatment [112]. To comprehend the mechanisms behind MT-enhanced antioxidant protection (guaiacol

peroxidase, GPX; GPX) maize seeding thermotolerance; catalase: CAT, ascorbic acid: ASA; and glutathione: GSS), detoxifying methylglyoxal (mg) (glyoxalase I: systems have been tests on Gly I; and Glyoxalase II: Gly II) and Osmoregulation (Proline: Pro; Trehalose, TSS) systems. The findings indicate that MT therapy promotes antioxidant (GPX, GR, CAT) and MG detoxification practices. Non-enzyme antioxidants (ASA and GSH) and osmolytes (Pro, Tre, and TSS) increased in enzymes (goly I and gly II), maize seedlings retained higher enzyme activity, and antioxidants under regular culture and the content of osmolyte is compared with the control under HS conditions. This study indicated that MT could improve the thermotolerance maize seedlings, MG detoxification, and Osmoregulation mechanisms through control of antioxidant response [113-117].

6. CONCLUSION

Melatonin has paramount importance in curbing the diseases caused by different fungi. Fungi cause most of the plant diseases, and recent researches has shown that melatonin has great importance in the prevention of fungal diseases in crop plants, and some of those researches are also discussed in this study. In recent years there has been a remarkable development in plant and melatonin research made. The improvement expands knowledge about the existence of melatonin, metabolism, and functions of seedlings. Melatonin is found in various types of organs and plants, as mentioned above, though precise concentrations are not stable in the various plants and organs. Biosynthetic pathway of melatonin is different in plants because there is no pineal gland in plants. The mechanism of melatonin synthesis in plants is similar to that of auxin mechanism in animals. Much researches have shown that melatonin has great importance in increasing resistance in plants against biotic and abiotic stresses. Endogenous concentrations in plants, melatonin rose under various stress conditions, from which we can conclude that melatonin is involved in enhancing tolerance in plants against different stresses. Different critical points related to melatonin like its metabolism pathway and its regulation because of the stress environment are still unknown.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Lerner AB, Case JD, Takahashi Y, Lee TH, Mori W. Isolation of melatonin, the pineal gland factor that lightens melanocyteS1. *J. Am. Soc.* 1958;80(10):2587-2597.
2. Lerner AB, Case JD. Part III: General Considerations of Skin Pigmentation: Pigment Cell Regulatory Factors. *J. Investig. Dermatol.* 1959;32(2):211-221.
3. Nordlund J.J, Lerner, A.B. The effects of oral melatonin on skin color and on the release of pituitary hormones. *J. Clin. Endocrinol. Metab.* 1977;45(4):768-774.
4. Reiter RJ. Melatonin: the chemical expression of darkness. *Mol. Cell. Endocrinol.* 1991;79(1-3):153-158.
5. Pelham RW. A serum melatonin rhythm in chickens and its abolition by pinealectomy. *Endocrinology.* 1975;96(2):543-546.
6. Yu HS, Pang SF, Tang PL, Brown GM. Persistence of circadian rhythms of melatonin and N-acetylserotonin in the serum of rats after pinealectomy. *Neuroendocrinology.* 1981;32(5):262-265.
7. Pevet P, Challet E. Melatonin: both master clock output and internal time-giver in the circadian clocks network. *J. Physiol.* 2011;105(4-6):170-182.
8. Vriend J, Reiter RJ. Melatonin feedback on clock genes: a theory involving the proteasome. *J. Pineal. Res.* 2015;58(1):1-9.
9. Legros C, Chesneau D, Boutin JA, Barc C, Malpoux B. Melatonin from cerebrospinal fluid but not from blood reaches sheep cerebral tissues under physiological conditions. *J. Neuroendocrinol.* 2014;26(3):151-163.
10. Reiter RJ, Tan DX, Kim SJ, Cruz MH. Delivery of pineal melatonin to the brain and SCN: role of canaliculi, cerebrospinal fluid, tanycytes and Virchow–Robin perivascular spaces. *Brain Struct. Funct.* 2014;219(6):1873-1887.
11. von Gall C, Stehle JH, Weaver DR. Mammalian melatonin receptors: molecular biology and signal transduction. *Cell Tiss. Res.* 2002;309(1):151-162.
12. Weaver DR, Reppert SM. The Mel1a melatonin receptor gene is expressed in human suprachiasmatic nuclei. *Neuroreport.* 1996;8(1):109-112.
13. Reiter RJ, Rosales-Corral S, Coto-Montes A, Boga JA, Tan DX, Davis JM, Konturek PC, Konturek SJ, Brzozowski T. Review

- atricle. *J. Physiol Pharmacol.* 2011; 62(3):269-274.
14. Pevet P, Agez L, Bothorel B, Saboureau M, Gauer F, Laurent V, Masson-Pévet M. Melatonin in the multi-oscillatory mammalian circadian world. *Chronobiol. Int.* 2006;23(1-2):39-51.
 15. Stehle JH, Von Gall C, Korf HW. Melatonin: a clock-output, a clock-input. *J. Neuroendocrinol.* 2003;15(4):383-389.
 16. Manchester LC, Poeggeler B, Alvares FL, Ogden GB, Reiter RJ. Melatonin immunoreactivity in the photosynthetic prokaryote *Rhodospirillum rubrum*: implications for an ancient antioxidant system. *Cell. Mol. Biol. Res.* 1995;41(5):391-395.
 17. Tilden, A.R.; Becker, M.A.; Amma, L.L.; Arciniega, J.; McGaw, A.K. Melatonin production in an aerobic photosynthetic bacterium: An evolutionarily early association with darkness. *J. Pineal Res.* 1997;22:102–106.
 18. Roopin M, Yacobi YZ, Levy O. Occurrence, diel patterns, and the influence of melatonin on the photosynthetic performance of cultured *Symbiodinium*. *J. Pineal Res.* 2013;55(1):89-100.
 19. Tan DX, Hardeland R, Manchester LC, Paredes SD, Korkmaz A, Sainz RM, Mayo JC, Fuentes-Broto L, Reiter RJ. The changing biological roles of melatonin during evolution: from an antioxidant to signals of darkness, sexual selection and fitness. *Biol. Rev.* 2010;85(3):607-623.
 20. Tosches MA, Bucher D, Vopalensky P, Arendt D. Melatonin signaling controls circadian swimming behavior in marine zooplankton. *Cell.* 2014;159(1):46-57.
 21. Wang F, Yeung KL, Chan WC, Kwok CC, Leung SL, Wu C, Chan EY, Yu IT, Yang XR, Tse LA. A meta-analysis on dose-response relationship between night shift work and the risk of breast cancer. *Ann. Oncol.* 2013;24(11):2724-2732.
 22. Åkerstedt T, Knutsson A, Narusyte J, Svedberg P, Kecklund G, Alexanderson K. Night work and breast cancer in women: a Swedish cohort study. *BMJ Open.* 2015;5(4):8127-8135.
 23. Papantoniou K, Castaño-Vinyals G, Espinosa A, Aragonés N, Pérez-Gómez B, Burgos J, Gómez-Acebo I, Llorca J, Peiró R, Jimenez-Moleón JJ, Arredondo F. Night shift work, chronotype and prostate cancer risk in the MCC-S pain case-control study. *Int. J. cancer.* 2015;137(5):1147-1157.
 24. Mirick DK, Bhatti P, Chen C, Nordt F, Stanczyk FZ, Davis S. Night shift work and levels of 6-sulfatoxymelatonin and cortisol in men. *Cancer Epidemiol Biomark. Prev.* 2013;22(6):1079-1087.
 25. Rehman RS, Zafar SA, Ali M, Pasha AN, Naveed MS, Waseem M, Raza A. CRISPR-Cas Mediated Genome Editing: A Paradigm Shift towards Sustainable Agriculture and Biotechnology. *Asian P. Res. J.* 2022;9(1):27-49.
 26. Rehman RS, Pasha AN, Zafar SA, Ali M, Waseem M, Ahmad M, Ahmad N, Hafeez AH. Chromosomal Engineering through CRISPR/Cas Technology: A Way Forward. *J. Adv. Bio. Biotech.* 2022;25(1):34-45.
 27. Stevens RG, Blask DE, Brainard GC, Hansen J, Lockley SW, Provencio I, Rea MS, Reinlib L. Meeting report: the role of environmental lighting and circadian disruption in cancer and other diseases. *Environ. Health Perspect.* 2007;115(9):1357-1362.
 28. Rehman RS, Ali M, Zafar SA, Ahmad M, Pasha AN, Bashir H, Rashid F, Hussain M. Tapping into the Unsung Potential of CRISPR/CAS Technology in Agriculture. *Asian J. Biochem. Gen. Mol. Bio.* 2022;10(4): 1-26.
 29. Reiter RJ. The melatonin rhythm: both a clock and a calendar. *Experientia.* 1993;49(8):654-664.
 30. Ralph CL, Harlow HJ, Phillips JA. Delayed effect of pinealectomy on hibernation of the golden-mantled ground squirrel. *Int. J. Biometeorol.* 1982;26(4):311-328.
 31. Reiter RJ. Influence of pinealectomy on the breeding capability of hamsters maintained under natural photoperiodic and temperature conditions. *Neuroendocrinology.* 1973;13(6):366-370.
 32. Byeon Y, Park S, Kim YS, Park DH, Lee S, Back K. Light-regulated melatonin biosynthesis in rice during the senescence process in detached leaves. *J. Pineal Res.* 2012;53(1):107-111.
 33. Van Tassel DL, Roberts N, Lewy A, O'Neill SD. Melatonin in plant organs. *J. Pineal Res.* 2001;31(1):8-15.
 34. Okazaki M, Ezura H. Profiling of melatonin in the model tomato (*Solanum lycopersicum* L.) cultivar Micro-Tom. *J. Pineal Res.* 2009;46(3):338-343.
 35. Shi H, Reiter RJ, Tan DX, Chan Z. INDOLE-3-ACETIC ACID INDUCIBLE 17

- positively modulates natural leaf senescence through melatonin-mediated pathway in *Arabidopsis*. J. Pineal Res. 2015;58(1):26-33.
36. Afreen F, Zobayed SM, Kozai T. Melatonin in *Glycyrrhiza uralensis*: response of plant roots to spectral quality of light and UV-B radiation. J. Pineal Res. 2006;41(2):108-115.
 37. Arnao MB, Hernández-Ruiz J. Growth conditions determine different melatonin levels in *Lupinus albus* L. J. Pineal Res. 2013 Sep;55(2):149-155.
 38. Rehman RS, Ali M, Zafar SA, Hussain M, Pasha A, Naveed MS, Ahmad M, Waseem M. Abscisic Acid Mediated Abiotic Stress Tolerance in Plants. Asian J. Res. C. Sci. 2022;7(1):1-17.
 39. Posmyk MM, Janas KM. Melatonin in plants. Acta Physiol. Plant. 2009;31(1):1-1.
 40. Murch SJ, KrishnaRaj S, Saxena PK. Tryptophan is a precursor for melatonin and serotonin biosynthesis in in vitro regenerated *St. John's wort* (*Hypericum perforatum* L. cv. Anthos) plants. Plant Cell Rep. 2000;19(7):698-704.
 41. Rehman RS, Zafar SA, Ali M, Pasha AN, Bashir H, Ashraf MA, Yaqoob MU, Hussain M. Biochemical and Molecular Insights into Abiotic Stress Tolerance in Plants. Asian J. Biotech. Gen. Engg. 2022;5(2):1-19.
 42. Tan DX, Hardeland R, Back K, Manchester LC, Alatorre-Jimenez MA, Reiter RJ. On the significance of an alternate pathway of melatonin synthesis via 5-methoxytryptamine: comparisons across species. J. Pineal Res. 2016;61(1):27-40.
 43. Choi GH, Lee HY, Back K. Chloroplast overexpression of rice caffeic acid O-methyltransferase increases melatonin production in chloroplasts via the 5-methoxytryptamine pathway in transgenic rice plants. J. Pineal Res. 2017;63(1):12412-12420.
 44. Rehman RS, Khan K, Zafar SA, Pasha AN, Ali M, Ashraf MA, Bashir H, Rashid F. Character Association Studies in Various *Brassica napus* Genotypes under Drought Stress. Asian J. Res. Bot. 2022;7(2):20-27.
 45. Byeon Y, Lee K, Park YI, Park S, Back K. Molecular cloning and functional analysis of serotonin N-acetyltransferase from the *Cyanobacterium synechocystis* sp. PCC 6803. J. Pineal Res. 2013;55(4):371-376.
 46. Semak I, Korik E, Antonova M, Wortsman J, Slominski A. Metabolism of melatonin by cytochrome P450s in rat liver mitochondria and microsomes. J. Pineal Res. 2008;45(4):515-523.
 47. Dibrova DV, Cherepanov DA, Galperin MY, Skulachev VP, Mulkidjanian AY. Evolution of cytochrome bc complexes: from membrane-anchored dehydrogenases of ancient bacteria to triggers of apoptosis in vertebrates. Biochim. Biophys. Acta. 2013;1827(11-12):1407-1427.
 48. Tesoriere L, Avellone G, Ceraulo L, D'Arpa D, Allegra M, Livrea MA. Oxidation of melatonin by oxoferryl hemoglobin: a mechanistic study. Free Radic. Res. 2001;35(6):633-642.
 49. Tan DX, Hardeland R, Manchester LC, Galano A, Reiter RJ. Cyclic-3-hydroxymelatonin (C3HOM), a potent antioxidant, scavenges free radicals and suppresses oxidative reactions. Curr. Med. Chem. 2014;21(13):1557-1565.
 50. Hardeland R, Tan DX, Reiter RJ. Kynuramines, metabolites of melatonin and other indoles: the resurrection of an almost forgotten class of biogenic amines. J. Pineal Res. 2009;47(2):109-126.
 51. Kołodziejczyk I, Bałabusta M, Szewczyk R, Posmyk MM. The levels of melatonin and its metabolites in conditioned corn (*Zea mays* L.) and cucumber (*Cucumis sativus* L.) seeds during storage. Acta Physiol. Plant. 2015;37(6):1-1.
 52. Byeon Y, Tan DX, Reiter RJ, Back K. Predominance of 2-hydroxymelatonin over melatonin in plants. J. Pineal Res. 2015;59(4):448-454.
 53. Byeon Y, Back K. Molecular cloning of melatonin 2-hydroxylase responsible for 2-hydroxymelatonin production in rice (*Oryza sativa*). J. Pineal Res. 2015;58(3):343-351.
 54. Tan DX, Hardeland R, Manchester LC, Poeggeler B, Lopez-Burillo S, Mayo JC, Sainz RM, Reiter RJ. Mechanistic and comparative studies of melatonin and classic antioxidants in terms of their interactions with the ABTS cation radical. J. Pineal Res. 2003;34(4):249-259.
 55. Lowes DA, Webster NR, Murphy MP, Galley HF. Antioxidants that protect mitochondria reduce interleukin-6 and oxidative stress, improve mitochondrial function, and reduce biochemical markers of organ dysfunction in a rat model of acute sepsis. Br. J. Anaesth. 2013;110(3):472-480.

56. Gitto E, Tan DX, Reiter RJ, Karbownik M, Manchester LC, Cuzzocrea S, Fulia F, Barberi I. Individual and synergistic antioxidative actions of melatonin: studies with vitamin E, vitamin C, glutathione and desferrioxamine (desferoxamine) in rat liver homogenates. *J. Pharm. Pharmacol.* 2001;53(10):1393-1401.
57. Sharma S, Haldar C. Melatonin prevents X-ray irradiation induced oxidative damage in peripheral blood and spleen of the seasonally breeding rodent, *Funambulus pennanti* during the reproductively active phase. *Int. J. Radiat. Biol.* 2006;82(6):411-419.
58. Túnez I, Munoz MC, Medina FJ, Salcedo M, Feijóo M, Montilla P. Comparison of melatonin, vitamin E and L-carnitine in the treatment of neuro- and hepatotoxicity induced by thioacetamide. *Cell Biochem. Funct.* 2007;25(2):119-127.
59. Hardeland R. Antioxidative protection by melatonin. *Endocrine.* 2005;27(2):119-130.
60. Hardeland R. Ubiquitous melatonin presence and effects in unicells, plants and animals. *Trends Comp. Biochem. Physiol.* 1996;2:25-45.
61. Hardeland R, Poeggeler B. Non-vertebrate melatonin. *J. Pineal Res.* 2003;34(4):233-241.
62. Hardeland R, Coto-Montes A, Poeggeler B. Circadian rhythms, oxidative stress, and antioxidative defense mechanisms. *Chronobiol. Int.* 2003;20(6):921-962.
63. Hardeland R, Pandi-Perumal SR. Melatonin, a potent agent in antioxidative defense: actions as a natural food constituent, gastrointestinal factor, drug and prodrug. *Nutr. Meta.* 2005;2(1):1-5.
64. Hsu CH, Han BC, Liu MY, Yeh CY, Casida JE. Phosphine-induced oxidative damage in rats: attenuation by melatonin. *Free Radic. Biol. Med.* 2000;28(4):636-642.
65. Guenther AL, Schmidt SI, Laatsch H, Fotso S, Ness H, Ressmeyer AR, Poeggeler B, Hardeland R. Reactions of the melatonin metabolite AMK (N1-acetyl-5-methoxykynuramine) with reactive nitrogen species: formation of novel compounds, 3-acetamidomethyl-6-methoxycinnolinone and 3-nitro-AMK. *J. Pineal Res.* 2005;39(3):251-260.
66. Johnston JD, Bashforth R, Diack A, Andersson H, Lincoln GA, Hazlerigg DG. Rhythmic melatonin secretion does not correlate with the expression of arylalkylamine N-acetyltransferase, inducible cyclic amp early repressor, period1 or cryptochrome1 mRNA in the sheep pineal. *Neuroscience.* 2004;124(4):789-795.
67. Jou MJ, Peng TI, Reiter RJ, Jou SB, Wu HY, Wen ST. Visualization of the antioxidative effects of melatonin at the mitochondrial level during oxidative stress-induced apoptosis of rat brain astrocytes. *J. Pineal Res.* 2004;37(1):55-70.
68. Klein DC, Weller JL. Indole metabolism in the pineal gland: a circadian rhythm in N-acetyltransferase. *Science.* 1970;169(3950):1093-1095.
69. Vázquez J, Grillitsch K, Daum G, Mas A, Torija MJ, Beltran G. Melatonin minimizes the impact of oxidative stress induced by hydrogen peroxide in *Saccharomyces* and non-conventional yeast. *Front. Microbiol.* 2018:1933-1945.
70. Costa V, Moradas-Ferreira P. Oxidative stress and signal transduction in *Saccharomyces cerevisiae*: insights into aging, apoptosis and diseases. *Mol. Aspects Med.* 2001;22(4-5):217-246.
71. Blask DE, Sauer LA, Dauchy RT, Holowachuk EW, Ruhoff MS, Kopff HS. Melatonin inhibition of cancer growth in vivo involves suppression of tumor fatty acid metabolism via melatonin receptor-mediated signal transduction events. *Cancer Res.* 1999;59(18):4693-4701.
72. Chen X, Sun C, Laborda P, Zhao Y, Palmer I, Fu ZQ, Qiu J, Liu F. Melatonin treatment inhibits the growth of *Xanthomonas oryzae* pv. *oryzae*. *Front. Microbiol.* 2018:2280-2286.
73. Blask DE, Sauer LA, Dauchy RT, Holowachuk EW, Ruhoff MS, Kopff HS. Melatonin inhibition of cancer growth in vivo involves suppression of tumor fatty acid metabolism via melatonin receptor-mediated signal transduction events. *Cancer Res.* 1999;59(18):4693-4701.
74. Bubis M, Zisapel N. Facilitation and inhibition of G-protein regulated protein secretion by melatonin. *Neurochem. Int.* 1995;27(2):177-183.
75. Byeon Y, Back K. Low melatonin production by suppression of either serotonin N-acetyltransferase or N-acetylserotonin methyltransferase in rice causes seedling growth retardation with yield penalty, abiotic stress susceptibility, and enhanced coleoptile growth under

- anoxic conditions. *J. Pineal Res.* 2016;60(3):348-359.
76. Hardeland R. Ubiquitous melatonin-presence and effects in unicells, plants and animals. *Trends Comp. Biochem. Physiol.* 1996;2:25-45.
 77. Reiter RJ, Tan DX, Burkhardt S, Manchester LC. Melatonin in plants. *Nutr. Rev.* 2001;59(9):286-290.
 78. Hardeland R. Melatonin and 5-methoxytryptamine in non-metazoans. *Reprod. Nutr. Dev.* 1999;39(3):399-408.
 79. Hardeland R, Poeggeler B. Non-vertebrate melatonin. *J. Pineal Res.* 2003;34(4):233-241.
 80. Hardeland R, Pandi-Perumal SR, Poeggeler B. Melatonin in plants: focus on a vertebrate night hormone with cytoprotective properties. *Funct. Plant Sci. Biotechnol.* 2007;1(1):32-45.
 81. Fuhrberg B, Hardeland R, Poeggeler B, Behrmann C. Dramatic rises of melatonin and 5-methoxytryptamine in *Gonyaulax* exposed to decreased temperature. *Biol. Rhythm Res.* 1997;28(1):144-150.
 82. Fuhrberg B, Hardeland R. Temperature as a major environmental factor controlling levels and rhythm amplitudes of melatonin in the marine dinoflagellate *Gonyaulax polyedra*. *Biometeorology.* 1997;14:272-277.
 83. Pandi-Perumal SR, Srinivasan V, Maestroni GJM, Cardinali DP, Poeggeler B, Hardeland R. Melatonin – Nature's most versatile biological signal? *FEBS J.* 2006; 273: 2813-2838.
 84. Poeggeler B, Balzer I, Hardeland R, Lerchl A. Pineal hormone melatonin oscillates also in the dinoflagellate *Gonyaulax polyedra*. *Naturwissenschaften.* 1991;78: 268-269.
 85. Poeggeler B, Hardeland R. Detection and quantification of melatonin in a dinoflagellate, *Gonyaulax polyedra*: Solutions to the problem of methoxyindole destruction in non-vertebrate material. *J. Pineal Res.* 1994;17(1):1-10.
 86. Rodriguez C, Mayo JC, Sainz RM, Antolín I, Herrera F, Martín V, Reiter RJ. Regulation of antioxidant enzymes: a significant role for melatonin. *J. Pineal Res.* 2004;36(1):1-9.
 87. Mueller U, Hardeland R, Fuhrberg B, Poeggeler B. Accumulation and metabolism of 5-methoxylated indoleamines in the dinoflagellate *Gonyaulax polyedra*. *Eur. J. Cell Biol.* 2000;79-85.
 88. Mueller U, Hardeland R, Poeggeler B, Fuhrberg B, Burkhardt S. Pathways of melatonin catabolism in the dinoflagellate *Gonyaulax polyedra*. *Biol. Rhythm Res.* 2001;32:465-470.
 89. Rehman RS, Pasha AN, Zafar SA, Ali M, Bashir H, Saeed MU, Ashraf NA, Javed A. Molecular Mechanisms behind the Regulation of Rice Tiller Angle: An Update. *Asian J. Biol.* 2022;14(4):37-50.
 90. Tan DX. Melatonin: a potent, endogenous hydroxyl radical scavenger. *Endocr j.* 1993;1:57-60.
 91. Tan DX, Hardeland R, Manchester LC, Poeggeler B, Lopez-Burillo S, Mayo JC, Sainz RM, Reiter RJ. Mechanistic and comparative studies of melatonin and classic antioxidants in terms of their interactions with the ABTS cation radical. *J. Pineal Res.* 2003;34(4):249-259.
 92. Sprenger J, Hardeland R, Fuhrberg B, Han SZ. Melatonin and other 5-methoxylated indoles in yeast: presence in high concentrations and dependence on tryptophan availability. *Cytologia.* 1999;64(2):209-213.
 93. Murch SJ, Simmons CB. Melatonin in feverfew and other medicinal plants. *Lancet.* 1997;350(9091):1598-1599.
 94. Murch SJ, Saxena PK. A melatonin-rich germplasm line of St John's wort (*Hypericum perforatum* L.). *J. Pineal Res.* 2006;41(3):284-287.
 95. Manchester LC, Tan DX, Reiter RJ, Park W, Monis K, Qi W. High levels of melatonin in the seeds of edible plants: possible function in germ tissue protection. *Life Sci.* 2000;67(25):3023-3029.
 96. Hardeland R, Pandi-Perumal SR, Poeggeler B. Melatonin in plants: focus on a vertebrate night hormone with cytoprotective properties. *Funct. Plant Sci. Biotechnol.* 2007;1(1):32-45.
 97. Chen G, Huo Y, Tan DX, Liang Z, Zhang W, Zhang Y. Melatonin in Chinese medicinal herbs. *Life Sci.* 2003;73(1):19-26.
 98. Hardeland R, Pandi-Perumal SR. Melatonin, a potent agent in antioxidative defense: actions as a natural food constituent, gastrointestinal factor, drug and prodrug. *Nutr. Meta.* 2005;2(1):1-5.
 99. Pandi-Perumal SR, Srinivasan V, Maestroni GJM, Cardinali DP, Poeggeler B, Hardeland R. Melatonin – Nature's most

- versatile biological signal? FEBS J. 2006; 273: 2813-2838.
100. Hardeland R, Poeggeler B. Actions of melatonin, its structural and functional analogs in the central nervous system and the significance of metabolism. *Cent. Nerv. Syst. Agents Med. Chem.* 2007;7(4):289-303.
 101. Reiter RJ, Tan DX, Manchester LC, Paredes SD, Mayo JC, Sainz RM. Melatonin and reproduction revisited. *Biol. Reprod.* 2009;81(3):445-456.
 102. Galano A, Tan DX, Reiter RJ. Melatonin as a natural ally against oxidative stress: a physicochemical examination. *J. Pineal Res.* 2011;51(1):1-6.
 103. Escames G, Ozturk G, Baño-Otálora B, Pozo MJ, Madrid JA, Reiter RJ, Serrano E, Concepción M, Acuña-Castroviejo D. Exercise and melatonin in humans: reciprocal benefits. *J. Pineal Res.* 2012;52(1):1-10.
 104. Venegas C, García JA, Escames G, Ortiz F, López A, Doerrier C, García-Corzo L, López LC, Reiter RJ, Acuña-Castroviejo D. Extrpineal melatonin: analysis of its subcellular distribution and daily fluctuations. *J. Pineal Res.* 2012;52(2):217-227.
 105. Xu W, Cai SY, Zhang Y, Wang Y, Ahammed GJ, Xia XJ, Shi K, Zhou YH, Yu JQ, Reiter RJ, Zhou J. Melatonin enhances thermotolerance by promoting cellular protein protection in tomato plants. *J. Pineal Res.* 2016;61(4):457-469.
 106. Wilson RA, Sangha MK, Banga SS, Atwal AK, Gupta S. Heat stress tolerance in relation to oxidative stress and antioxidants in *Brassica juncea*. *J. Environ. Biol.* 2014; 35(2):383-395.
 107. Salvucci ME, Crafts-Brandner SJ. Inhibition of photosynthesis by heat stress: the activation state of Rubisco as a limiting factor in photosynthesis. *Physiol. Plant.* 2004;120(2):179-186.
 108. Lazar D, Murch SJ, Beilby MJ, Al Khazaaly S. Exogenous melatonin affects photosynthesis in characeae *Chara australis*. *Plant Signal Behav.* 2013;8(3):23279-23295.
 109. Cheng F, Yin LL, Zhou J, Xia XJ, Shi K, Yu JQ, Zhou YH, Foyer CH. Interactions between 2-Cys peroxiredoxins and ascorbate in autophagosome formation during the heat stress response in *Solanum lycopersicum*. *J. Exp. Bot.* 2016;67(6):1919-1933.
 110. Pieterse CM, Leon-Reyes A, Van der Ent S, Van Wees SC. Networking by small-molecule hormones in plant immunity. *Nat. Chem. Biol.* 2009;5(5):308-316.
 111. Mittler R, Vanderauwera S, Gollery M, Van Breusegem F. Reactive oxygen gene network of plants. *Trends Plant Sci.* 2004;9(10):490-498.
 112. Li ZG, Xu Y, Bai LK, Zhang SY, Wang Y. Melatonin enhances thermotolerance of maize seedlings (*Zea mays* L.) by modulating antioxidant defense, methylglyoxal detoxification, and osmoregulation systems. *Protoplasma.* 2019;256(2):471-490.
 113. Fan J, Xie Y, Zhang Z, Chen L. Melatonin: a multifunctional factor in plants. *Int. J. Mol. Sci.* 2018;19(5):1528-1535.
 114. Gill SS, Tuteja N. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol. Biochem.* 2010;48(12):909-930.
 115. Hasanuzzaman M, Nahar K, Alam M, Roychowdhury R, Fujita M. Physiological, biochemical, and molecular mechanisms of heat stress tolerance in plants. *Int. J. Mol. Sci.* 2013;14(5):9643-3684.
 116. Hasanuzzaman M, Nahar K, Hossain M, Mahmud JA, Rahman A, Inafuku M, Oku H, Fujita M. Coordinated actions of glyoxalase and antioxidant defense systems in conferring abiotic stress tolerance in plants. *Int. J. Mol. Sci.* 2017; 18(1):200-215.
 117. Rehman RS, Zafar SA, Ali M, Ahmad M, Pasha AN, Waseem M, Hafeez AH, Raza A. Plant Pan-genomes: A new frontier in understanding genomic diversity in plants. *J. Adv. Bio. Biotech.* 2022;25(1):10-22.

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